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This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53 (c)

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TITLE OF THE INVENTION (280 characters max)

HETEROLOGOUS G-CSF FUSION PROTEINS

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ENCLOSED APPLICATION PARTS (check all that apply)

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The invention was made by an agency of the United States Government or under a contract with an agency of the United States Government.

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SIGNATURE 

Date 03/05/02

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43,936

 Additional inventors are being named on separately numbered sheets attached hereto

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## HETEROLOGOUS G-CSF FUSION PROTEINS

## FIELD OF THE INVENTION

The present invention relates to heterologous fusion proteins, including analogs and derivatives thereof, fused to proteins that have the effect of extending the *in vivo* half-life of the proteins. These fusion proteins are significant in human medicine, particularly in the treatment of conditions treatable by stimulation of circulating neutrophils, such as after chemotherapy regimens or in chronic congenital neutropenia. More specifically, the invention relates to novel heterologous fusion proteins with granulocyte-colony stimulating factor activity.

15

BACKGROUND OF THE INVENTION

Among all blood cell lineages, the modulation of neutrophil and platelet production has been of highest interest to clinical oncologists and hematologists. Myelosuppression is the single most severe complication of cancer chemotherapy, and a major cause of treatment delay during multiple-cycle or combination chemotherapy. It is also the major dose-limiting factor for most chemotherapeutic agents. Due to the short half-lives of neutrophils in peripheral blood, life-threatening falls in

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neutrophil levels are seen after a number of conventional anti-tumor chemotherapy regimens.

5 The most prominent regulator of granulopoiesis is granulocyte-colony stimulating factor (G-CSF). G-CSF induces proliferation and differentiation of hematopoietic progenitor cells resulting in increased numbers of circulating neutrophils. G-CSF also stimulates the release of mature neutrophils from bone marrow and activates their 10 functional state. [Souza L.M., et al. (1986) *Science* 232:61-65]. Thus, therapeutic proteins with G-CSF activity have tremendous value in situations where there are reduced 15 circulating levels of neutrophilic granulocytes.

15 However, the usefulness of therapy using G-CSF peptides has been limited by their short plasma half-life. Thus, they must be administered intravenously or subcutaneously at fairly frequent intervals (once or twice a day) in order to maintain their neutrophil stimulating properties. In 20 addition, this short half-life limits the performance of the drug to traditional drug delivery systems. It would clearly benefit the treatment of patients with abnormally low neutrophils, and reduce the discomfort and inconvenience associated with frequent injections to provide a pharmaceutical agent that could be administered less 25 frequently and optionally by alternative routes of administration. Thus, a need exists to develop agents that stimulate the production of mature neutrophils and are more optimal in their duration of effect.

30 The present invention overcomes the problems associated with delivering a compound that has a short plasma half-life in two respects. First, G-CSF is hyperglycosylated. The carbohydrate content of G-CSF is altered by substituting amino acids that can act as substrates for glycosylating enzymes in mammalian cells. Most significantly, the present 35 invention encompasses fusion of these hyperglycosylated G-

CSF analogs to another protein with a long circulating half-life such as the Fc portion of an immunoglobulin or albumin.

BRIEF SUMMARY OF THE INVENTION

5 Compounds of the present invention include heterologous fusion proteins comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of

10 a) human albumin;  
b) human albumin analogs; and  
c) fragments of human albumin.

15 Additional compounds of the present invention include a heterologous fusion protein comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of

20 a) the Fc portion of an immunoglobulin;  
b) an analog of the Fc portion of an immunoglobulin;  
and  
c) fragments of the Fc portion of an immunoglobulin.

25 The hyperglycosylated G-CSF analog may be fused to the polypeptide via a peptide linker. It is preferable that the peptide linker is selected from the group consisting of:

a) a glycine rich peptide;  
b) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>n</sub> where n is 1, 2, 3, 4, or 5; and  
c) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>3</sub>.

30 One aspect of the present invention includes heterologous fusion proteins, wherein the hyperglycosylated G-CSF analogs have the Formula (I) [SEQ ID NO:1]

1 5 10 15  
Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys  
20 25 30  
35 Xaa Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln  
35 40 45  
Glu Lys Leu Cys Xaa Xaa Xaa Lys Leu Cys His Pro Glu Glu Leu Val  
50 55 60  
Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa

		65	70	75	80											
	Xaa	Xaa	Xaa	Xaa	Gln	Leu	Ala	Gly	Cys	Leu	Ser	Gln	Leu	His	Ser	
					85				90					'35		
5	Gly	Leu	Phe	Leu	Tyr	Gln	Gly	Leu	Leu	Gln	Ala	Leu	Xaa	Xaa	Ser	
					100				105					110		
	Xaa	Glu	Leu	Gly	Pro	Thr	Leu	Asp	Thr	Leu	Gln	Leu	Asp	Val	Ala	Asp
					115				120					125		
	Phe	Ala	Thr	Thr	Ile	Trp	Gln	Gln	Met	Glu	Glu	Leu	Gly	Met	Ala	Pro
					130				135					140		
10	Ala	Leu	Gln	Pro	Xaa	Xaa	Xaa	Ala	Met	Pro	Ala	Phe	Xaa	Xaa	Xaa	Phe
					145				150					155		
	Gln	Arg	Arg	Ala	Gly	Gly	Val	Leu	Val	Ala	Ser	His	Leu	Gln	Ser	Phe
					165				170							
	Leu	Glu	Val	Ser	Tyr	Arg	Val	Leu	Arg	His	Leu	Ala	Gln	Pro		(I)

15 wherein:

Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;

Xaa at position 37 is Ala or Asn;

Xaa at position 38 is Thr, or any other amino acid except

20 Pro;

Xaa at position 39 is Tyr, Thr, or Ser;

Xaa at position 57 is Pro or Val;

Xaa at position 58 is Trp or Asn;

Xaa at position 59 is Ala or any other amino acid except

25 Pro;

Xaa at position 60 is Pro, Thr, Asn, or Ser,

Xaa at position 61 is Leu, or any other amino acid except

Pro;

Xaa at position 62 is Ser or Thr;

30 Xaa at position 63 is Ser or Asn;

Xaa at position 64 is Cys or any other amino acid except

Pro;

Xaa at position 65 is Pro, Ser, or Thr;

Xaa at position 66 is Ser or Thr;

35 Xaa at position 67 is Gln or Asn;

Xaa at position 68 is Ala or any other amino acid except

Pro;

Xaa at position 69 is Leu, Thr, or Ser

Xaa at position 93 is Glu or Asn

40 Xaa at position 94 is Gly or any other amino acid except

Pro;

Xaa at position 95 is Ile, Asn, Ser, or Thr;

Xaa at position 97 is Pro, Ser, Thr, or Asn;

Xaa at position 133 is Thr or Asn;

45 Xaa at position 134 is Gln or any other amino acid except

Pro;

Xaa at position 135 is Gly, Ser, or Thr

Xaa at position 141 is Ala or Asn;

Xaa at position 142 is Ser or any other amino acid except

50 Pro; and

Xaa at position 143 is Ala, Ser, or Thr;

and wherein:

5        Xaa at positions 37, 38, and 39 constitute region 1;  
Xaa at positions 58, 59, and 60 constitute region 2;  
Xaa at positions 59, 60, and 61 constitute region 3;  
Xaa at positions 60, 61, and 62 constitute region 4;  
Xaa at positions 61, 62, and 63 constitute region 5;  
Xaa at positions 62, 63, and 64 constitute region 6;  
10      Xaa at positions 63, 64, and 65 constitute region 7;  
Xaa at positions 64, 65, and 66 constitute region 8;  
Xaa at positions 67, 68, and 69 constitute region 9;  
Xaa at positions 93, 94, and 95 constitute region 10;  
Xaa at positions 94, 95, and Ser at position 96  
15      constitute region 11;  
Xaa at positions 95, and 97, and Ser at position 96  
constitute region 12;  
Xaa at positions 133, 134, and 135 constitute  
region 13;  
20      Xaa at positions 141, 142, and 143 constitute  
region 14;

and provided that at least one of regions 1 through 14  
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any  
25 amino acid except Pro and Xaa2 is Ser or Thr.

Thus, the heterologous fusion proteins of the present  
invention include analogs wherein one or any combination of  
two or more regions comprise the sequence Asn Xaa1 Xaa2  
wherein Xaa1 is any amino acid except Pro and Xaa2 is Ser or  
30 Thr.

Preferred hyperglycosylated G-CSF analogs that make up  
part of the heterologous fusion proteins of the present  
invention, include the following:

35      a) G-CSF[A37N, Y39T]

- b) G-CSF [P57V, W58N, P60T]
- c) G-CSF [P60N, S62T]
- d) G-CSF [S63N, P65T]
- e) G-CSF [Q67N, L69T]
- 5 f) G-CSF [E93N, I95T]
- g) G-CSF [T133N, G135T]
- h) G-CSF [A141N, A143T]
- i) G-CSF [A37N, Y39T, P57V, W58N, P60T]
- j) G-CSF [A37N, Y39T, P60N, S62T]
- 10 k) G-CSF [A37N, Y39T, S63N, P65T]
- l) G-CSF [A37N, Y39T, Q67N, L69T]
- m) G-CSF [A37N, Y39T, E93N, I95T]
- n) G-CSF [A37N, Y39T, T133N, G135T]
- o) G-CSF [A37N, Y39T, A141N, A143T]
- 15 p) G-CSF [A37N, Y39T, P57V, W58N, P60T, S63N, P65T]
- q) G-CSF [A37N, Y39T, P57V, W58N, P60T, Q67N, L69T]
- r) G-CSF [A37N, Y39T, S63N, P65T, E93N, I95T]

The present invention also includes heterologous fusion proteins, which are the product of the expression in a host cell of an exogenous DNA sequence, which comprises a DNA sequence encoding a heterologous fusion protein of Formula I (described above) fused to a DNA sequence encoding human albumin or the Fc portion of an immunoglobulin.

A hyperglycosylated heterologous fusion protein of the present invention also includes polynucleotides encoding the heterologous fusion protein described herein, vectors comprising these polynucleotides and host cells transfected or transformed with the vectors described herein. Also included is a process for producing a heterologous fusion protein comprising the steps of transcribing and translating a polynucleotide described herein under conditions wherein the heterologous fusion protein is expressed in detectable amounts.

The present invention encompasses a method for increasing neutrophil levels in a mammal comprising the administration of a therapeutically effective amount of a heterologous fusion protein described above. The present 5 invention also includes the use of the heterologous fusion proteins described above for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

The present invention also encompasses a pharmaceutical 10 formulation adapted for the treatment of patients with insufficient neutrophil levels comprising a glycosylated protein as described above.

#### BRIEF DESCRIPTION OF THE DRAWINGS

15 The invention is further illustrated with reference to the following drawings:

Figure 1: Schematic illustrating fourteen regions in human G-CSF wherein the amino acid sequence can be mutated to create functional glycosylation sites.

#### DETAILED DESCRIPTION OF THE INVENTION

The present invention comprises heterologous fusion proteins. As used herein, the term heterologous fusion protein means a hyperglycosylated G-CSF analog fused to 25 human albumin, a human albumin analog, a human albumin fragment, the Fc portion of an immunoglobulin, an analog of the Fc portion of an immunoglobulin, or a fragment of the Fc portion of an immunoglobulin. The G-CSF analog may be fused directly, or fused via a peptide linker, to an albumin or Fc protein. The albumin and Fc portion may be fused to the G- 30 CSF analogs at either terminus or at both termini. These heterologous fusion proteins are biologically active and have an increased half-life compared to native G-CSF.

*Hyperglycosylated G-CSF Analogs*

Hyperglycosylated analogs of G-CSF as used herein refers to analogs of G-CSF with one or more changes in the amino acid sequence which result in an increase in the number of sites for carbohydrate attachment compared with native human G-CSF expressed in animal cells *in vivo*. In addition, hyperglycosylated G-CSF analogs include human G-CSF wherein the O-linked glycosylation site at position 133 is replaced with an N-linked glycosylation site. Analogs are generated by site directed mutagenesis having substitution of amino acid residues creating new sites that are available for glycosylation. G-CSF analogs having a greater carbohydrate content than that found in native human G-CSF are generated by adding glycosylation sites that do not perturb the secondary, tertiary, and quaternary structure required for activity. The heterologous fusion proteins of the present invention thus have a larger mass and an increased negative charge compared to native G-CSF, they will not be as rapidly cleared from the circulation.

It is preferred that the hyperglycosylated G-CSF analog portion of the heterologous fusion protein have 1, 2, 3, or 4 additional sites for N-glycosylation. Figure 1 illustrates fourteen different regions that can be glycosylated with very little effect on *in vitro* activity.

Each region may be mutated to the consensus site for N-glycosylation addition which is Asn X1 X2 wherein X1 is any amino acid except Pro and X2 is Ser or Thr. It is preferred that the X1 amino acid be any other amino acid except Trp, Asp, Glu, or Leu and it is most preferred that the X1 amino acid be the naturally occurring amino acid. The scope of the present invention includes analogs wherein a single region (1 through 14) is mutated or wherein a region is mutated in combination with one or more other regions.

A representative number of fusion proteins having single or multiple additional glycosylation sites have been expressed, purified, and characterized. The mutated amino

acids in the hyperglycosylated G-CSF portion of the heterologous fusion protein are identified in parentheses. For example, G-CSF[A37N,Y39T] is G-CSF wherein the amino acids at positions 37 and 39 have been substituted to create 5 a glycosylation site. This site of carbohydrate attachment is illustrated as region 1 in Figure 1. G-CSF[A37N,Y39T,P57V,W58N,P60T] is an example of a G-CSF analog wherein amino acids in region 1 and region 2 are mutated to provide two functional glycosylation sites on a 10 single molecule (Figure 1).

G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T] is an example of a G-CSF analog wherein the amino acids in region 1, region 2, and region 9 are mutated to provide three functional glycosylation sites on a single molecule (Figure 15 1).

Native G-CSF can be used as the backbone to create the hyperglycosylated G-CSF analog portion of a particular heterologous fusion protein. In addition, the native G-CSF backbone can be modified such that substitutions in the 20 regions defined in Figure 1 are made in the context of a different or improved heterologous fusion protein. For example, native G-CSF with a Cysteine to Alanine substitution at position 17 may reduce aggregation and enhance stability and thus, can be used as the backbone used 25 to create the hyperglycosylated G-CSF analog portion of the heterologous fusion proteins of the present invention.

In addition, Reidhaar-Olson, et al., through alanine scanning mutagenesis, describe residues critical to the activity of human G-CSF. [Reidhaar-Olson, et al. (1996) 30 *Biochemistry* 35:9034-9041; See also Young, et al. (1997) *Protein Science* 6:1228-1236]. Thus, the hyperglycosylated G-CSF analogs can be modified by substituting amino acids outside the glycosylated regions described in Figure 1.

The heterologous fusion proteins of the present invention also encompass G-CSF analogs wherein the O-linked glycosylation site at position 133 is mutated to serve as an N-linked glycosylation site. The N-linked carbohydrate will 5 generally have a higher sialic acid content which will protect it from the rapid clearance mechanisms associated with native G-CSF.

The functions of a carbohydrate chain greatly depend on the structure of the attached carbohydrate moiety.

10 Typically compounds with a higher sialic acid content will have better stability and longer half-lives *in vivo*. The N-linked oligosaccharides contain sialic acid in both an  $\alpha$ 2,3 and an  $\alpha$ 2,6 linkage to galactose. [Takeuchi, et al. (1988) *J. Biol. Chem.* 263:3657]. Typically the sialic acid in the 15  $\alpha$ 2,3 linkage is added to galactose on the mannose  $\alpha$ 1,6 branch and the sialic acid in the  $\alpha$ 2,6 linkage is added to the galactose on the mannose  $\alpha$ 1,3 branch. The enzymes that add these sialic acids ( $\beta$ -galactoside  $\alpha$ 2,3 sialyltransferase and  $\beta$ -galactoside  $\alpha$ 2,6 sialyltransferase) are most efficient 20 at adding sialic acid to the mannose  $\alpha$ 1,6 and mannose  $\alpha$ 1,3 branches respectively.

Tetra-antennary N-linked oligosaccharides most commonly provide four possible sites for sialic acid attachment while bi- and tri-antennary oligosaccharide chains, which can 25 substitute for the tetra-antennary form at Asn-linked sites, commonly have at most only two or three sialic acids attached. O-linked oligosaccharides commonly provide only two sites for sialic acid attachment. Mammalian cell cultures can be screened for those cells that preferentially 30 add teta-antennary chains to the G-CSF analogs of the present invention, thereby maximizing the number of sites for sialic acid attachment. Different types of mammalian cells also differ with respect to the transferase enzymes present and consequently the sialic acid content and type of 35 oligosaccharide attached at each site. One way to optimize the carbohydrate content for a given G-CSF analog is to

express the analog in a cell line wherein an expression plasmid containing DNA encoding a specific sialyl transferase (e.g.,  $\alpha$ 2,6 sialyltransferase) is co-transfected with the G-CSF analog expression plasmid.

5 Alternatively a host cell line may be stably transfected with a sialyl transferase cDNA and that host cell used to express the G-CSF analog of interest. Thus, it is preferable if the oligosaccharide structure and sialic acid content are optimized for each analog encompassed by the  
10 present invention.

Heterologous Fc fusion proteins:

The hyperglycosylated G-CSF analogs described above can be fused directly or via a peptide linker to the Fc portion 15 of an immunoglobulin. For example SEQ ID NO:23 represents the sequence of a G-CSF-IgG1 Fc fusion protein, SEQ ID NO:24 represents the sequence of a G-CSF-IgG4 Fc fusion protein, and SEQ ID NO:25 represents the sequence of a G-CSF-human  
20 albumin fusion protein.

Immunoglobulins are molecules containing polypeptide chains held together by disulfide bonds typically having two light chains and two heavy chains. In each chain, one domain (V) has a variable amino acid sequence depending on the antibody specificity of the molecule. The other domains 25 (C) have a rather constant sequence common to molecules of the same class.

As used herein, the Fc portion of an immunoglobulin has the meaning commonly given to the term in the field of immunology. Specifically, this term refers to an antibody 30 fragment which is obtained by removing the two antigen binding regions (the Fab fragments) from the antibody. Thus, the Fc portion is formed from approximately equal sized fragments of the constant region from both heavy chains, which associate through non-covalent interactions 35 and disulfide bonds. The Fc portion can include the hinge regions and extend through the CH2 and CH3 domains to the C-

terminus of the antibody. For example, SEQ ID NOS: 18 and 19 represent the sequence of a Fc region encompassing the hinge region as well as the CH2 and CH3 domains. Other representative hinge regions for human and mouse

5 immunoglobulins can be found in Antibody Engineering, A Practical Guide, Borrebaeck, C.A.K., ed., W.H. Freeman and Co., 1992, the teachings of which are herein incorporated by reference.

There are five types of human immunoglobulin Fc regions  
10 with different effector and pharmacokinetic properties: IgG, IgA, IgM, IgD, and IgE. IgG is the most abundant immunoglobulin in serum. IgG also has the longest half-life in serum of any immunoglobulin (23 days). Unlike other immunoglobulins, IgG is efficiently recirculated following  
15 binding to an Fc receptor. There are four IgG subclasses G1, G2, G3, and G4, each of which have different effector functions. G1, G2, and G3 can bind C1q and fix complement while G4 cannot. Even though G3 is able to bind C1q more efficiently than G1, G1 is more effective at mediating complement-directed cell lysis. G2 fixes complement very  
20 inefficiently. The C1q binding site in IgG is located at the carboxy terminal region of the CH2 domain.

All IgG subclasses are capable of binding to Fc receptors (CD16, CD32, CD64) with G1 and G3 being more effective than G2 and G4. The Fc receptor-binding region of IgG is formed by residues located in both the hinge and the carboxy terminal regions of the CH2 domain.

IgA can exist both in a monomeric and dimeric form held together by a J-chain. IgA is the second most abundant Ig in serum, but it has a half-life of only 6 days. IgA has three effector functions. It binds to an IgA specific receptor on macrophages and eosinophils, which drives phagocytosis and degranulation, respectively. It can also fix complement via an unknown alternative pathway.

35 IgM is expressed as either a pentamer or a hexamer, both of which are held together by a J-chain. IgM has a

serum half-life of 5 days. It binds weakly to C1q via a binding site located in its CH3 domain. IgD has a half-life of 3 days in serum. It is unclear what effector functions are attributable to this Ig. IgE is a monomeric Ig and has 5 a serum half-life of 2.5 days. IgE binds to two Fc receptors which drives degranulation and results in the release of pro-inflammatory agents.

Depending on the desired *in vivo* effect, the heterologous fusion proteins of the present invention may 10 contain any of the isotypes described above or may contain mutated Fc regions wherein the complement and/or Fc receptor binding functions have been altered. For example, one embodiment of the present invention is a heterologous fusion protein wherein the Fc portion comprises a human IgG4 sequence wherein serine at position 229 is changed to 15 proline which reduces the effector function of the protein. (See SEQ ID NO: 22).

The heterologous fusion proteins of the present invention may contain the entire Fc portion of an 20 immunoglobulin, fragments of the Fc portion of an immunoglobulin, or analogs thereof fused to a G-CSF analog. Furthermore, the Fc portion may be fused at either terminus or at both termini.

The heterologous fusion proteins of the present 25 invention can consist of single chain proteins or as multi-chain polypeptides. Two or more Fc fusion proteins can be produced such that they interact through disulfide bonds that naturally form between Fc regions. These multimers can be homogeneous with respect to the G-CSF analog or they may 30 contain different G-CSF analogs fused at the N-terminus of the Fc portion of the fusion protein.

Regardless of the final structure of the fusion protein, the Fc or Fc-like region serves to prolong the *in vivo* plasma half-life of the G-CSF analog compared to native 35 G-CSF. Furthermore, the fused G-CSF analog must retain some

biological activity. Biological activity can be determined by in vitro and in vivo methods known in the art.

Since the Fc region of IgG produced by proteolysis has the same in vivo half-life as the intact IgG molecule and

5 Fab fragments are rapidly degraded, it is believed that the relevant sequence for prolonging half-life resides in the CH2 and/or CH3 domains. Further, it has been shown in the literature that the catabolic rates of IgG variants that do not bind the high-affinity Fc receptor or C1q are  
10 indistinguishable from the rate of clearance of the parent wild-type antibody, indicating that the catabolic site is distinct from the sites involved in Fc receptor or C1q binding. [Wawrzynczak et al., (1992) *Molecular Immunology* 29:221]. Site-directed mutagenesis studies using a murine  
15 IgG1 Fc region suggested that the site of the IgG1 Fc region that controls the catabolic rate is located at the CH2-CH3 domain interface.

Based on these studies, Fc regions can be modified at the catabolic site to optimize the half-life of the fusion proteins. It is preferable that the Fc region used for the heterologous fusion proteins of the present invention be derived from an IgG1 such as that represented by SEQ ID NO: 21 or an IgG4 Fc region such as that represented by SEQ ID NO: 22. It is even more preferable that the Fc region be  
25 IgG4 or derived from IgG4. Preferably the IgG Fc region contains both the CH2 and CH3 regions including the hinge region.

Heterologous albumin fusion proteins:

30 The G-CSF analogs described above can be fused directly or via a peptide linker to albumin or an analog, fragment, or derivative thereof. SEQ ID NO:25 provides a representative G-CSF-human albumin fusion protein.

Generally the albumin proteins making up part of the  
35 heterologous fusion proteins of the present invention can be derived from albumin cloned from any species. However,

human albumin and fragments and analogs thereof are preferred to reduce the risk of the fusion protein being immunogenic in humans. Human serum albumin (HA) consists of a single non-glycosylated polypeptide chain of 585 amino

5 acids with a formula molecular weight of 66,500. The amino acid sequence of human albumin is represented as SEQ ID NO:

20. [See Meloun, et al. (1975) FEBS Letters 58:136; Behrens, et al. (1975) Fed. Proc. 34:591; Lawn, et al. (1981) Nucleic Acids Research 9:6102-6114; Minghetti, et al.

10 (1986) J. Biol. Chem. 261:6747]. A variety of polymorphic variants as well as analogs and fragments of albumin have been described. [See Weitkamp, et al., (1973) Ann. Hum. Genet. 37:219]. For example, in EP 322,094, the inventors disclose various shorter forms of HA. Some of these

15 fragments include HA(1-373), HA(1-388), HA(1-389), HA(1-369), and HA(1-419) and fragments between 1-369 and 1-419. EP 399,666 discloses albumin fragments that include HA(1-177) and HA(1-200) and fragments between HA(1-177) and HA(1-200).

20 It is understood that the heterologous fusion proteins of the present invention include hyperglycosylated G-CSF analogs that are coupled to any albumin protein including fragments, analogs, and derivatives wherein such fusion protein is biologically active and has a longer plasma half-life than native G-CSF. Thus, the albumin portion of the fusion protein need not necessarily have a plasma half-life equal to that of native human albumin. In addition, the albumin may be fused to either terminus or both termini of the hyperglycosylated G-CSF analog.

25 The heterologous fusion proteins of the present invention encompass proteins having conservative amino acid substitutions in the hyperglycosylated G-CSF analog and/or the Fc or albumin portion of the fusion protein.

30 The term "amino acid" is used herein in its broadest sense, and includes naturally occurring amino acids as well as non-naturally occurring amino acids, including amino acid

analog and derivatives. The latter includes molecules containing an amino acid moiety. One skilled in the art will recognize, in view of this broad definition, that reference herein to an amino acid includes, for example, 5 naturally occurring proteogenic L-amino acids; D-amino acids; chemically modified amino acids such as amino acid analogs and derivatives; naturally occurring non-proteogenic amino acids such as norleucine,  $\beta$ -alanine, ornithine, GABA, etc.; and chemically synthesized compounds having properties 10 known in the art to be characteristic of amino acids. As used herein, the term "proteogenic" indicates that the amino acid can be incorporated into a peptide, polypeptide, or protein in a cell through a metabolic pathway.

The incorporation of non-natural amino acids, including 15 synthetic non-native amino acids, substituted amino acids, or one or more D-amino acids into the heterologous fusion proteins of the present invention can be advantageous in a number of different ways. D-amino acid-containing peptides, etc., exhibit increased stability *in vitro* or *in vivo* 20 compared to L-amino acid-containing counterparts. Thus, the construction of peptides, etc., incorporating D-amino acids can be particularly useful when greater intracellular stability is desired or required. More specifically, D-peptides, etc., are resistant to endogenous peptidases and 25 proteases, thereby providing improved bioavailability of the molecule, and prolonged lifetimes *in vivo* when such properties are desirable. Additionally, D-peptides, etc., cannot be processed efficiently for major histocompatibility complex class II-restricted presentation to T helper cells, 30 and are therefore less likely to induce humoral immune responses in the whole organism.

Although the heterologous fusion proteins of the present invention can be made by a variety of different methods, recombinant methods are preferred.

*Construction of DNA encoding the heterologous fusion proteins of the present invention:*

Wild type albumin and immunoglobulin proteins can be obtained from a variety of sources. For example, these 5 proteins can be obtained from a cDNA library prepared from tissue or cells which express the mRNA of interest at a detectable level. Libraries can be screened with probes designed using the published DNA or protein sequence for the particular protein of interest.

10 Screening a cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1989). An alternative means to isolate a gene encoding an 15 albumin or immunoglobulin protein is to use PCR methodology [Sambrook et al., *supra*; Dieffenbach et al., *PCR Primer: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1995)]. PCR primers can be designed based on published sequences.

20 Generally the full-length wild-type sequences cloned from a particular species can serve as a template to create analogs, fragments, and derivatives that retain the ability to confer a longer plasma half-life on the G-CSF analog that is part of the fusion protein. It is preferred that the Fc 25 and albumin portions of the heterologous fusion proteins of the present invention be derived from the native human sequence in order to reduce the risk of potential immunogenicity of the fusion protein in humans.

30 In particular, it is preferred that the immunoglobulin portion of a fusion protein encompassed by the present invention contain only an Fc fragment of the immunoglobulin. Depending on whether particular effector functions are desired and the structural characteristics of the fusion protein, an Fc fragment may contain the hinge region along 35 with the CH2 and CH3 domains or some other combination thereof. These Fc fragments can be generated using PCR

techniques with primers designed to hybridize to sequences corresponding to the desired ends of the fragment.

Similarly, if fragments of albumin are desired, PCR primers can be designed which are complementary to internal albumin sequences. PCR primers can also be designed to create restriction enzyme sites to facilitate cloning into expression vectors.

DNA encoding human G-CSF can be obtained from a cDNA library prepared from tissue or cells which express G-CSF mRNA at a detectable level such as monocytes, macrophages, vascular endothelial cells, fibroblasts, and some human malignant and leukemic myeloblastic cells. Libraries can be screened with probes designed using the published DNA sequence for human G-CSF. [Souza L., et al. (1986) *Science* 232:61-65]. Screening a cDNA or genomic library with the selected probe may be conducted using standard procedures, such as described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1989). An alternative means to isolate the gene encoding human G-CSF is to use PCR methodology [Sambrook, et al., *supra*; Dieffenbach, et al., *PCR Primer: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, NY (1995)].

The glycosylated G-CSF analogs of the present invention can be constructed by a variety of mutagenesis techniques well known in the art. Specifically, a representative number of glycosylated G-CSF analogs were constructed using mutagenic PCR from a cloned wild-type human G-CSF DNA template (Example 1).

The glycosylated G-CSF analogs of the present invention may be produced by other methods including recombinant DNA technology or well known chemical procedures, such as solution or solid-phase peptide synthesis, or semi-synthesis in solution beginning with protein fragments coupled through conventional solution methods.

Recombinant DNA methods are preferred for producing the glycosylated G-CSF analogs of the present invention. Host

cells are transfected or transformed with expression or cloning vectors described herein for glycosylated G-CSF analog production and cultured in conventional nutrient media modified as appropriate for inducing promoters, 5 selecting transformants, or amplifying the genes encoding the desired sequences (Example 2). The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation

Physical stability is an essential feature for 10 therapeutic formulations. The physical stability of the heterologous fusion proteins of the present invention depends on their conformational stability, the number of charged residues (pI of the protein), the ionic strength and pH of the formulation, and the protein concentration, among 15 other possible factors. As discussed previously, the G-CSF analog portion of the heterologous fusion proteins can be successfully glycosylated and expressed such that the three dimensional structure is maintained. Because these heterologous fusion proteins are able to fold properly in a 20 hyperglycosylated state, they will have improved conformational and physical stability relative to wild-type G-CSF.

While wild-type G-CSF produced in mammalian cells and bacterial cells has similar activity *in vivo*, the mammalian 25 cell-produced protein has increased conformational and physical stability due to the presence of a single O-linked sugar moiety present at position 133. Thus, the G-CSF analog portion of the heterologous fusion proteins, which has an increased glycosylation content compared to wild-type G- 30 CSF produced in mammalian or bacterial cells, will confer increased stability on the heterologous fusion protein. Furthermore, it is likely that glycosylation may inhibit inter-domain interactions and consequently enhance stability by preventing inter-domain disulfide shuffling.

35 The gene encoding a heterologous fusion protein can be constructed by ligating DNA encoding a G-CSF analog in-frame

to DNA encoding an albumin or Fc protein. The gene encoding the G-CSF analog and the gene encoding the albumin or Fc protein can also be joined in-frame via DNA encoding a linker peptide.

5 The in vivo function and stability of the heterologous fusion proteins of the present invention can be optimized by adding small peptide linkers to prevent potentially unwanted domain interactions. Although these linkers can potentially be any length and consist of any combination of amino acids,  
10 it is preferred that the length be no longer than necessary to prevent unwanted domain interactions and/or optimize biological activity and/or stability. Generally, the linkers should not contain amino acids with extremely bulky side chains or amino acids likely to introduce significant  
15 secondary structure. It is preferred that the linker be serine-glycine rich and be less than 30 amino acids in length. It is more preferred that the linker be no more than 20 amino acids in length. It is even more preferred that the linker be no more than 15 amino acids in length. A  
20 preferred linker contains repeats of the sequence Gly-Gly-Gly-Gly-Ser. It is preferred that there be between 2 and 6 repeats of this sequence. It is even more preferred that there be between 3 and 4 repeats of this sequence.

To construct the heterologous G-CSF fusion proteins, the  
25 DNA encoding wild-type G-CSF, albumin, and Fc polypeptides and fragments thereof can be mutated either before ligation or in the context of a cDNA encoding an entire fusion protein. A variety of mutagenesis techniques are well known in the art. For example, a mutagenic PCR method utilizes  
30 strand overlap extension to create specific base mutations for the purposes of changing a specific amino acid sequence in the corresponding protein. This PCR mutagenesis requires the use of four primers, two in the forward orientation (primers A and C) and two in the reverse orientation  
35 (primers B and D). A mutated gene is amplified from the wild-type template in two different stages. The first

reaction amplifies the gene in halves by performing an A to B reaction and a separate C to D reaction wherein the B and C primers target the area of the gene to be mutated. When aligning these primers with the target area, they contain 5 mismatches for the bases that are targeted to be changed. Once the A to B and C to D reactions are complete, the reaction products are isolated and mixed for use as the template for the A to D reaction. This reaction then yields the full, mutated product.

10 Once a gene encoding an entire fusion protein is produced it can be cloned into an appropriate expression vector. Specific strategies that can be employed to make the G-CSF fusion proteins of the present invention are described in example 1.

15 *General methods to recombinantly express the heterologous fusion proteins of the present invention:*

Host cells are transfected or transformed with expression or cloning vectors described herein for heterologous fusion 20 protein production and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences. The culture conditions, such as media, temperature, pH and the like, can be selected by the skilled artisan without undue experimentation. In general, principles, protocols, and practical techniques for maximizing the productivity of cell cultures can be found in *Mammalian Cell Biotechnology: A Practical Approach*, M. Butler, ed. (IRL Press, 1991) and Sambrook, et al., *supra*. Methods of 25 transfection are known to the ordinarily skilled artisan, for example, CaPO<sub>4</sub> and electroporation. General aspects of mammalian cell host system transformations have been described in U.S. Patent No. 4,399,216. Transformations into yeast are typically carried out according to the method of van Solingen, 30 et al., *J Bact.* 130(2): 946-7 (1977) and Hsiao, et al., *Proc. Natl. Acad. Sci. USA* 76(8): 3829-33 (1979). Suitable host 35

cells for the expression of the fusion proteins of the present invention are derived from multicellular organisms.

The fusion proteins of the present invention may be recombinantly produced directly, or as a protein having a signal sequence or other additional sequences which create a specific cleavage site at the N-terminus of the mature fusion protein. In general, the signal sequence may be a component of the vector, or it may be a part of the fusion protein-encoding DNA that is inserted into the vector. The signal sequence may be a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, lpp, or heat-stable enterotoxin II leaders. For yeast secretion the signal sequence may be, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces* cc-factor leaders, the latter described in U.S. Patent No. 5,010,182), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179), or the signal described in WO 90/13646. In mammalian cell expression, mammalian signal sequences may be used to direct secretion of the protein, such as signal sequences from secreted polypeptides of the same or related species as well as viral secretory leaders.

An example of suitable selectable markers for mammalian cells are those that enable the identification of cells competent to take up the fusion protein-encoding nucleic acid, such as DHFR or thymidine kinase. An appropriate host cell when wild-type DHFR is employed is the CHO cell line deficient in DHFR activity, prepared and propagated as described [Urlaub and Chasin, *Proc. Natl. Acad. Sci. USA*, 77(7): 4216-20 (1980)]. A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid Yrp7 [Stinchcomb, et al., *Nature* 282(5734): 39-43 (1979); Kingsman, et al., *Gene* 7(2): 141-52 (1979); Tschumper, et al., *Gene* 10(2): 157-66 (1980)]. The *trp1* gene provides a selection marker for a mutant strain of yeast lacking the ability to grow in

tryptophan, for example, ATCC No. 44076 or PEPC1 [Jones, *Genetics* 85: 23-33 (1977)].

Various forms of a fusion protein may be recovered from culture medium or from host cell lysates. If membrane-bound,  
5 it can be released from the membrane using a suitable detergent solution (e.g., Triton-X 100) or by enzymatic cleavage. Cells employed in expression of a fusion protein can be disrupted by various physical or chemical means, such as freeze-thaw cycling, sonication, mechanical disruption, or  
10 cell lysing agents.

*Purification of the heterologous fusion proteins of the present invention:*

Once the heterologous fusion proteins of the present invention are expressed in the appropriate host cell, the analogs can be isolated and purified. The following procedures are exemplary of suitable purification procedures:

Various methods of protein purification may be employed and such methods are known in the art and described, for example, in Deutscher, *Methods in Enzymology* 182: 83-9 (1990) and Scopes, *Protein Purification: Principles and Practice*, Springer-Verlag, NY (1982). The purification step(s) selected will depend on the nature of the production process used and the particular fusion protein produced. For example, fusion proteins comprising an Fc fragment can be effectively purified using a Protein A or Protein G affinity matrix. Low or high pH buffers can be used to elute the fusion protein from the affinity matrix. Mild elution conditions will aid in preventing irreversible denaturation of the fusion protein.  
25 Imidazole-containing buffers can also be used. Example 3 describes some successful purification protocols for the fusion proteins of the present invention.

*Characterization of the heterologous fusion proteins of the present invention:*

Numerous methods exist to characterize the fusion proteins of the present invention. Some of these methods include: SDS-PAGE coupled with protein staining methods or immunoblotting using anti-IgG, anti-HA and anti-G-CSF antibodies. Other methods include matrix assisted laser desorption/ionization-mass spectrometry (MALDI-MS), liquid chromatography/mass spectrometry, isoelectric focusing, analytical anion exchange, chromatofocussing, and circular dichroism to name a few. A representative number of heterologous fusion proteins were characterized using SDS-PAGE coupled with immunoblotting as well as mass spectrometry

For example, Table 2 illustrates the calculated molecular mass for a representative number of fusion proteins as well as the observed mass (as measured by protease mapping/LC-MS).

The relative differences between observed mass and mass calculated for a nonglycosylated protein are indicative of the extent of glycosylation.

Table 2

Construct	Theoretical mass (kDa)	Observed mass (kDa)
C17A (GCSF)-IgG1 Fc	89.4	93.0
C17A (GCSF)-IgG4 Fc	88.7	92.7
C17A (GCSF)-HSA	86.0	85.1
C17A, A37N, S63N, E93N (GCSF)-IgG1 Fc	89.4	97-114 (104.7)
C17A, A37N, S63N, E93N (GCSF)-IgG4 Fc	88.7	96-112 (103.8)
C17A, A37N, S63N, E93N (GCSF)-HSA	86.0	87-98 (92.5)
C17A, A37N, W58N, Q67N (GCSF)-IgG1 Fc	89.3	95-110 (103.8)
C17A, A37N, W58N, Q67N (GCSF)-IgG4 Fc	88.8	95-110 (102.4)
C17A, A37N, W58N, Q67N (GCSF)-HSA	85.9	87-98 (93.1)

*Administration and Preparation of Compositions:*

The heterologous fusion proteins of the present invention may be formulated with one or more excipients. The active

fusion proteins of the present invention may be combined with a pharmaceutically acceptable buffer, and the pH adjusted to provide acceptable stability, and a pH acceptable for administration such as parenteral administration.

5        Optionally, one or more pharmaceutically-acceptable antimicrobial agents may be added. Meta-cresol and phenol are preferred pharmaceutically-acceptable microbial agents. One or more pharmaceutically-acceptable salts may be added to adjust the ionic strength or tonicity. One or more excipients  
10      may be added to adjust the isotonicity of the formulation. Glycerin is an example of an isotonicity-adjusting excipient. Pharmaceutically acceptable means suitable for administration to a human or other animal and thus, does not contain toxic elements or undesirable contaminants and does not interfere  
15      with the activity of the active compounds therein.

      A pharmaceutically-acceptable salt form of the heterologous fusion proteins of the present invention may be used in the present invention. Acids commonly employed to form acid addition salts are inorganic acids such as hydrochloric acid, hydrobromic acid, hydriodic acid, sulfuric acid, phosphoric acid, and the like, and organic acids such as p-toluenesulfonic acid, methanesulfonic acid, oxalic acid, p-bromophenyl-sulfonic acid, carbonic acid, succinic acid, citric acid, benzoic acid, acetic acid, and the like.  
20      Preferred acid addition salts are those formed with mineral acids such as hydrochloric acid and hydrobromic acid.  
25      Preferred acid addition salts are those formed with mineral acids such as hydrochloric acid and hydrobromic acid.

      Base addition salts include those derived from inorganic bases, such as ammonium or alkali or alkaline earth metal hydroxides, carbonates, bicarbonates, and the like. Such bases useful in preparing the salts of this invention thus include sodium hydroxide, potassium hydroxide, ammonium hydroxide, potassium carbonate, and the like.  
30      Bases useful in preparing the salts of this invention thus include sodium hydroxide, potassium hydroxide, ammonium hydroxide, potassium carbonate, and the like.

      Administration may be via any route known to be effective  
35      by the physician of ordinary skill. Peripheral parenteral is one such method. Parenteral administration is commonly

understood in the medical literature as the injection of a dosage form into the body by a sterile syringe or some other mechanical device such as an infusion pump. Peripheral parenteral routes can include intravenous, intramuscular, 5 subcutaneous, and intraperitoneal routes of administration.

The heterologous fusion proteins of the present invention may also be amenable to administration by oral, rectal, nasal, or lower respiratory routes, which are non-parenteral routes. Of these non-parenteral routes, the lower respiratory route 10 and the oral route are preferred.

The heterologous fusion proteins of the present invention can be used to treat patients with insufficient circulating neutrophil levels, typically those undergoing cancer chemotherapy.

An "effective amount" of the heterologous fusion protein is the quantity which results in a desired therapeutic and/or prophylactic effect without causing unacceptable side-effects when administered to a subject in need of G-CSF receptor stimulation. A "desired therapeutic effect" includes one or more of the following: 1) an amelioration of the symptom(s) associated with the disease or condition; 2) a delay in the onset of symptoms associated with the disease or condition; 3) increased longevity compared with the absence of the treatment; and 4) greater 15 quality of life compared with the absence of the treatment. 20

The present invention comprises G-CSF compounds that have improved biochemical and biophysical properties by virtue of being fused to an albumin protein, an albumin fragment, an albumin analog, a Fc protein, a Fc fragment, or 25 a Fc analog. These heterologous proteins can be successfully expressed in host cells, retain signaling activities associated with activation of the G-CSF receptor, and have prolonged half-lives.

EXAMPLESExample 1: Construction of DNA encoding heterologous fusion proteins:

Table 1 provides the sequence of primers used to create functional glycosylation sites in different regions of the G-CSF protein (See Figure 1).

Table 1: Primer sequences used to introduce mutations into human G-CSF.

Mutation	A Primer*	B Primer*	C Primer*	D Primer*
WT	CF177 [SEQ ID NO:26] <b>GTAAGCTTGCCT</b> <b>CGACGCTAGCCTG</b> <b>CGCGCCGCCATG</b> GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	CF178 [SEQ ID NO:27] GGGGCAGGGAGC TGGCTGGGCCA GTGGAGTGGCTT CCTGCACTGTCC AGAGTGCACTGT G	CF179 [SEQ ID NO:28] GGACAGTGCAGG AAGCCACTCCAC TGGGCCAGCCA GCTCCCTGCC AGAGCTTCCTG AGAGCTTCCTG	CF176 [SEQ ID NO:29] <b>GAACCTCGAGGA</b> <b>TCCTCATTAGGG</b> <b>CTGGGCAAGGTG</b> CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
C17A <i>SacI</i>	CF177 [SEQ ID NO:30] <b>GTAAGCTTGCCT</b> <b>CGACGCTAGCCTG</b> <b>CGCGCCGCCATG</b> GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	C17Arev [SEQ ID NO:31] GCTCTAAGGCCT TGAGCAGGAAGC TCTGGGCAGGG AGCTCGCTGGC CCAGTGGAG	C17Afor [SEQ ID NO:32] GGGCCCAGCGAG CTCCCTGCC GAGCTTCCTGCT CAAGGCCTT GCAAG	CF176 [SEQ ID NO:33] <b>GAACCTCGAGGA</b> <b>TCCTCATTAGGG</b> <b>CTGGGCAAGGTG</b> CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
A37N, Y39T <i>SpeI</i>	CF177 [SEQ ID NO:34] <b>GTAAGCTTGCCT</b> <b>CGACGCTAGCCTG</b> <b>CGCGCCGCCATG</b> GCCGGACCTGCC ACCCAGAGCCCC ATGAAGCTG	A37Nrev [SEQ ID NO:35] GTCCGAGCAGCA CTAGTTCTCGG GGTGGCACAGCT TGGTGGTGT ACAGCTTCTC G	A37Nfor [SEQ ID NO:36] GGCGCAGCGCTC CAGGAGAACGCTG TGTAAACACC AAGCTGTGCCAC CCCGAGGA GTGCTG	CF176 [SEQ ID NO:37] <b>GAACCTCGAGGA</b> <b>TCCTCATTAGGG</b> <b>CTGGGCAAGGTG</b> CCTTAAGACGCG GTACGACACCTC CAGGAAGCTCTG
T133N, G135T	CF177 [SEQ ID NO:38]	T133Nrev [SEQ ID NO:39]	T133Nfor [SEQ ID NO:40]	CF176 [SEQ ID NO:41]

<i>Eco47III</i>	<b>GTAAGCTTGC GT</b> <b>CGACGCTAGCGG</b> <b>CGCGCCGCCATG</b> <b>GCCGGACCTGCC</b> <b>ACCCAGAGCCCC</b> <b>ATGAAGCTG</b>	<b>GCCCCGGCGCTGG</b> <b>AAAGCGCTGGCG</b> <b>AAGGCCGGCATG</b> <b>GCGGTCTGGTTG</b> <b>GGCTGCAGGGCA</b> <b>G</b>	<b>GGCCCCCTGCCCT</b> <b>GCAGCCCAACCA</b> <b>GACCGCCATGCC</b> <b>GGCCTTCGCCAG</b> <b>CGCTTCCAGCG</b>	<b>GAACCTCGAGGA</b> <b>TCCTCATTAGGG</b> <b>CTGGGCAAGGTG</b> <b>CCTTAAGACGCG</b> <b>GTACGACACCTC</b> <b>CAGGAAGCTCTG</b>
<i>A141N, A143T <i>SapI</i></i>	<b>CF177 [SEQ ID NO: 42]</b> <b>GTAAGCTTGC GT</b> <b>CGACGCTAGCGG</b> <b>CGCGCCGCCATG</b> <b>GCCGGACCTGCC</b> <b>ACCCAGAGCCCC</b> <b>ATGAAGCTG</b>	<b>A141Nrev [SEQ ID NO: 43]</b> <b>GCCCCGGCGCTGG</b> <b>AAGGTAGAGTTG</b> <b>AAGGCCGGCATG</b> <b>GCACCCCTGGGTG</b> <b>GGCTGAAGAGCA</b> <b>GGGGCCAT</b>	<b>A141Nfor [SEQ ID NO: 44]</b> <b>GGGAATGGCCCC</b> <b>TGCTCTTCAGCC</b> <b>CACCCAGGGTGC</b> <b>CATGCCGGCCTT</b> <b>CAACTCTACCTT</b> <b>CCAGCGCCGGGC</b>	<b>CF176 [SEQ ID NO: 45]</b> <b>GAACCTCGAGGA</b> <b>TCCTCATTAGGG</b> <b>CTGGGCAAGGTG</b> <b>CCTTAAGACGCG</b> <b>GTACGACACCTC</b> <b>CAGGAAGCTCTG</b>
<i>P57V, W58N, P60T <i>HpaI</i></i>	<b>JCB128 [SEQ ID NO: 46]</b> <b>GCTAGCGGCGCG</b> <b>CCACCATG</b>	<b>JCB136 [SEQ ID NO: 47]</b> <b>GCTCAGGGTAGC</b> <b>GTAAACGATGCC</b> <b>CAGAGAGTG</b>	<b>JCB137 [SEQ ID NO: 48]</b> <b>GGGCATCGTTAA</b> <b>CGCTACCCCTGAG</b> <b>CAGCTG</b>	<b>JCB129 [SEQ ID NO: 49]</b> <b>GACTCGAGGATC</b> <b>CTCATTAGGGCT</b> <b>GGG</b>
<i>Q67N, L69T <i>NaeI</i></i>	<b>JCB134 [SEQ ID NO: 50]</b> <b>GCTAGCGGCGCG</b> <b>CCACCATGGCCG</b> <b>GACCTGCCACCC</b> <b>AG</b>	<b>JCB138 [SEQ ID NO: 51]</b> <b>CAAGCAGCCGGC</b> <b>CAGCTGGGTGGC</b> <b>GTGCTGGGGCA</b> <b>GCTGCTCAG</b>	<b>JCB139 [SEQ ID NO: 52]</b> <b>GCCCCAGCAACG</b> <b>CCACCCAGCTGG</b> <b>CCGGCTGCTTGA</b> <b>G</b>	<b>JCB135 [SEQ ID NO: 53]</b> <b>GACTCGAGGATC</b> <b>CTCATTAGGGCT</b> <b>GGGCAAGGTGCC</b> <b>TTAAGACGCGG</b>
<i>P60N, S62T <i>SpeI</i></i>	<b>JCB128 [SEQ ID NO: 54]</b> <b>GCTAGCGGCGCG</b> <b>CCACCATG</b>	<b>JCB130 [SEQ ID NO: 55]</b> <b>GGGGCAACTAGT</b> <b>CAGGTTAGCCCA</b> <b>GGG</b>	<b>JCB131 [SEQ ID NO: 56]</b> <b>GCTAACCTGACT</b> <b>AGTTGCCCCAGC</b> <b>CAG</b>	<b>JCB129 [SEQ ID NO: 57]</b> <b>GACTCGAGGATC</b> <b>CTCATTAGGGCT</b> <b>GGG</b>
<i>S63N, P65T <i>MfeI</i></i>	<b>JCB128 [SEQ ID NO: 58]</b> <b>GCTAGCGGCGCG</b> <b>CCACCATG</b>	<b>JCB132 [SEQ ID NO: 59]</b> <b>GGTGCAATTGCT</b> <b>CAGGGGAGCCCA</b>	<b>JCB133 [SEQ ID NO: 60]</b> <b>GCAATTGCACCA</b> <b>GCCAGGCCCTG</b>	<b>JCB129 [SEQ ID NO: 61]</b> <b>GACTCGAGGATC</b> <b>CTCATTAGGGCT</b>

		G		GGG
E93N, I95T <i>BspEI</i>	JCB134 [SEQ ID NO:62] <b>GCTAGCGCGCG</b> <b>CCACCATGGCCG</b> <b>GACCTGCCACCC</b> AG	JCB140 [SEQ ID NO:63] <b>CCGGACTGGTCC</b> <b>CGTTCAGGCCT</b> <b>GCAGGAGCCCT</b> G	JCB141 [SEQ ID NO:64] <b>GAACGGGACCA</b> <b>TCCGGAGTTGGG</b> <b>TCCCACCTTGG</b>	JCB135 [SEQ ID NO:65] <b>GACTCGAGGATC</b> <b>CTCATTTAGGGCT</b> <b>GGGCAAGGTGCC</b> <b>TTAAGACGCCG</b>
<i>Sall</i>	JCB155 [SEQ ID NO:66] <b>GTTCGACGCTAGC</b> <b>GGCGCGCCACCA</b> <b>TGGCCGGACCTG</b>			

\*Nucleotides in bold represent changes imposed in the target sequence and nucleotides in bold and italics represent flanking sequences which may add restriction sites to facilitate cloning, Kozac sequences, or stop codons.

Preparation 1a: DNA encoding wild-type human G-CSF

A strand overlapping extension PCR reaction was used to create a wild type human G-CSF construct in order to eliminate the methylation of an *ApaI* site. Isolated human G-CSF cDNA served as the template for these reactions. The 5' end A primer was used to create a restriction enzyme site prior to the start of the coding region as well as to introduce a Kozac sequence (GGCGCC) 5' of the coding leader sequence to facilitate translation in cell culture.

The A-B product was generated using primers CF177 and CF178 in a PCR reaction. Likewise, the C-D product was produced with primers CF179 and CF176. The products were isolated and combined. The combined mixture was then used as a template with primers CF177 and CF178 to create the full-length wild-type construct. [Nelson, R.M. and Long, G.C. (1989), *Anal. Biochem.* 180:147-151].

The full-length product was ligated into the pCR2.1-Topo vector (Invitrogen, Inc. Cat. No. K4500-40) by way of a topoisomerase TA overhang system to create pCR2.1G-CSF.

The following protocol was used for preparation of the full-length wild-type G-CSF protein as well as each of the G-CSF analogs. Approximately 5 ng of template DNA and 15 pmol of each primer was used in the initial PCR reactions.

5 The reactions were prepared using Platinum PCR Supermix® (GibcoBRL Cat. No. 11306-016). The PCR reactions were denatured at 94°C for 5 min and then subject to 25 cycles wherein each cycle consisted of 30 seconds at 94°C followed by 30 seconds at 60°C followed by 30 seconds at 72°C. A 10 final extension was carried out for 7 minutes at 72°C. PCR fragments were isolated from agarose gels and purified using a Qiaquick® gel extraction kit (Qiagen, Cat. No. #28706). DNA was resuspended in sterile water and used for the final PCR reaction to prepare full-length product.

15 Preparation 1b: DNA encoding G-CSF[A37N, Y39T, P57V, W58N, P60T, Q67N, L69T] was constructed as follows:

DNA encoding G-CSF[A37N, Y39T, Q67N, L69T] was subcloned 20 into pJB02 to create pJB02G-CSF[A37N, Y39T, Q67N, L69T] and pJB02G-CSF[A37N, Y39T, P57V, W58N, P60T] served as the template for strand overlapping expression PCR. JCB155 and JCB136 served as the A and B primers and JCB137 and JCB135 served 25 as the C and D primers. The full-length mutated cDNA was prepared as described previously using JCB155 and JCB134 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1, region 2, and region 9 of the protein (See Figure 1). The full-length cDNA was ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N, Y39T, P57V, W58N, P60T, 30 Q67N, L69T].

Preparation 1c: DNA encoding G-CSF[A37N, Y39T, S63N, P64T, E93N, I95T] was constructed as follows:

DNA encoding G-CSF[A37N, Y39T, E93N, I95T] was subcloned 35 into pJB02 to create pJB02G-CSF[A37N, Y39T, E93N, I95T] and pJB02G-CSF[A37N, Y39T, E93N, I95T] served as the template for

strand overlapping expression PCR. JCB155 and JCB132 served as the A and B primers and JCB133 and JCB135 served as the C and D primers. The full-length mutated cDNA was prepared as described previously using JCB155 and JCB135 primers. The 5 resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1, region 7, and region 10 of the protein (See Figure 1). The full-length cDNA was ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N, Y39T, S63N, P64T, E93N, I95T].

10 Preparation 1d: DNA encoding G-CSF[C17A] which is G-CSF wherein the amino acid at position 17 is substituted with Ala is constructed as follows:

15 The wild-type construct in the pCR2.1-Topo vector (pCR2.1G-CSF) serves as the PCR template for the C17A mutagenesis. Strand overlapping extension PCR is performed as described previously. CF177 and C17Arev serve as the A-B primers and C17Afor and CF176 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously 20 using the CF177 and CF176 primers. The B and C primers are used to mutate the DNA such that a SacI restriction site is created and the protein expressed from the full-length sequence contains an Alanine instead of a Cysteine at position 17. The full-length cDNA is ligated back into the 25 pCR2.1-Topo vector to create pCR2.1G-CSF[C17A] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the Nhe/Xho sites of mammalian expression vector pJB02 to create pJB02G-CSF[C17A].

30 Preparation 1e: DNA encoding G-CSF[A37N, Y39T] is constructed as follows:

35 Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and A37Nrev serve as the A-B primers and CF176 and A37Nfor serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the CF177 and CF176 primers.

The B and C primers contain mismatched sequences such that a *SpeI* site is created in the DNA and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 1 of the protein. The 5 full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[A37N,Y39T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[A37N,Y39T].

10

Preparation 1f: DNA encoding G-CSF[P57V,W58N,P60T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB136 serve as the A-B primers and JCB137 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *HpaI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 2 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[P57V,W58N,P60T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe/Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[P57V,W58N,P60T].

Preparation 1g: DNA encoding G-CSF[P60N,S62T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB130 serve as the A-B primers and JCB131 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. 35 The B and C primers contain mismatched sequences such that a *SpeI* site is created and the protein expressed from the

full-length sequence contains a consensus sequence for N-linked glycosylation in region 4 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[P60N,S62T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe*/Xho sites of mammalian expression vector pJB02 to create pJB02G-CSF[P60N,S62T].

Preparation 1h: DNA encoding G-CSF[S63N,P65T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB128 and JCB132 serve as the A-B primers and JCB133 and JCB129 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *Mfe*I site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 7 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[S63N,P65T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe*/Xho sites of mammalian expression vector pJB02 to create pJB02G-CSF[S63N,P65T].

Preparation 1i: DNA encoding G-CSF[Q67N,L69T] is constructed as follows:

Strand overlapping extension PCR is performed using pJB02G-CSF[C17A] as the template. Primers JCB134 and JCB138 serve as the A-B primers and JCB139 and JCB135 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that a *Nae*I site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 9 of the protein. The full-

length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[Q67N,L69T] wherein the sequence is confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe*/*Xho* sites of mammalian expression vector pJB02 to create 5 pJB02G-CSF[Q67N,L69T].

Preparation 1j: DNA encoding G-CSF[E93N,I95T] is constructed as follows:

Strand overlapping extension PCR is performed using 10 pJB02G-CSF[C17A] as the template. Primers JCB134 and JCB140 serve as the A-B primers and JCB141 and JCB135 serve as the C-D primers. The full-length mutated cDNA is prepared as described previously using the JCB128 and JCB129 primers. The B and C primers contain mismatched sequences such that 15 a *Bsp*EI site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 10 of the protein. The full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[E93N,I95T] wherein the sequence is 20 confirmed. G-CSF analog encoding DNA is then cloned into the *Nhe*/*Xho* sites of mammalian expression vector pJB02 to create pJB02G-CSF[E93T,I95T].

Preparation 1k: DNA encoding G-CSF[T133N,G135T] is 25 constructed as follows:

Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and T133Nrev serve as the A-B primers and T133Nfor and CF176 serve as the C-D primers. The full-length mutated cDNA is 30 prepared as described previously using the CF177 and CF176 primers. The B and C primers contain mismatched sequences such that an *Eco*47III site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 13 of the 35 protein. The full-length cDNA is ligated back into the

pCR2.1-Topo vector to create pCR2.1G-CSF[T133N, G135T] wherein the sequence is confirmed.

Preparation 11: DNA encoding G-CSF[A141N, A143T] is  
5 constructed as follows:

Strand overlapping extension PCR is performed using pCR2.1G-CSF[C17A] as the template. Primers CF177 and A141Nrev serve as the A-B primers and A141Nfor and CF176 serve as the C-D primers. The full-length mutated cDNA is 10 prepared as described previously using the CF177 and CF176 primers. The B and C primers contain mismatched sequences such that an *SapI* site is created and the protein expressed from the full-length sequence contains a consensus sequence for N-linked glycosylation in region 14 of the protein. The 15 full-length cDNA is ligated back into the pCR2.1-Topo vector to create pCR2.1G-CSF[A141N, A143T] wherein the sequence is confirmed.

Preparation 1m: DNA encoding G-CSF[A37N, Y39T, T133N, G135T] is constructed as follows:

A 210 bp insert containing G-CSF[A37N, Y39T] is isolated from pCR2.1G-CSF[A37N, Y39T] using *EcoNI*. This fragment is ligated into pCR2.1G-CSF[T133N, G135T] which is prepared by cleavage with *EcoNI* and subsequent isolation of the vector 25 (4359 bp) from a 210 bp fragment containing wild-type G-CSF sequences. This ligation creates pCR2.1G-CSF[A37N, Y39T, T133N, G135T]. Analog encoding DNA is then subcloned into pJB02 using *NheI/XhoI* to create pJB02G-CSF[A37N, Y39T, T133N, G135T].

30 Preparation 1n: DNA encoding G-CSF[A37N, Y39T, A141N, A143T] is constructed as follows:

A 210 bp insert containing G-CSF[A37N, Y39T] is isolated from pCR2.1G-CSF[A37N, Y39T] using *EcoNI*. This fragment is 35 ligated into pCR2.1G-CSF[A141N, A143T] which is prepared by cleavage with *EcoNI* and subsequent isolation of the vector

(4359 bp) from a 210 bp fragment containing wild-type G-CSF sequences. This ligation creates pCR2.1G-CSF[A37N, Y39T, A141N, A143T]. Analog encoding DNA is then subcloned into pJB02 using *Nhe*I/*Xho*I to create pJB02G-CSF[A37N, Y39T, A141N, A143T].

5 Preparation 1o: DNA encoding G-CSF[A37N, Y39T, P57V, W58N, P60T] is constructed as follows:

DNA encoding G-CSF[A37N, Y39T] is subcloned into pJB02 to create pJB02G-CSF[A37N, Y39T] and pJB02G-CSF[A37N, Y39T] serves as the template for strand overlapping expression PCR. JCB128 and JCB136 serve as the A and B primers and JCB137 and JCB129 serve as the C and D primers. The full-length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 2 of the protein. The full-length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N, Y39T, P57V, W58N, P60T].

20 Preparation 1p: DNA encoding G-CSF[A37N, Y39T, Q67N, L69T] is constructed as follows:

DNA encoding G-CSF[A37N, Y39T] is subcloned into pJB02 to create pJB02G-CSF[A37N, Y39T] and pJB02G-CSF[A37N, Y39T] serves as the template for strand overlapping expression PCR. JCB134 and JCB138 serve as the A and B primers and JCB139 and JCB135 serve as the C and D primers. The full-length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 9 of the protein. The full-length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF[A37N, Y39T, Q67N, L69T].

35 Preparation 1q: DNA encoding G-CSF[A37N, Y39T, E93N, I95T] is constructed as follows:

DNA encoding G-CSF [A37N, Y39T] is subcloned into pJB02 to create pJB02G-CSF [A37N, Y39T] and pJB02G-CSF [A37N, Y39T] serves as the template for strand overlapping expression PCR. JCB134 and JCB140 serve as the A and B primers and 5 JCB141 and JCB135 serve as the C and D primers. The full-length mutated cDNA is prepared as described previously using JCB128 and JCB129 primers. The resulting full-length DNA encodes a protein with consensus N-linked glycosylation sites in region 1 and region 10 of the protein. The full- 10 length cDNA is ligated back into pCR2.1-Topo to create pCR2.1G-CSF [A37N, Y39T, E93N, I95T].

Various mutated G-CSF polypeptides were then ligated in-frame directly to DNA encoding IgG1 or IgG4. Various 15 mutated G-CSF polypeptides were also ligated in-frame via DNA encoding a peptide linker or directly to human albumin.

Example 2: Expression of heterologous fusion proteins:

20 2a: Expression in 293/EBNA cells:

Each full-length DNA encoding a heterologous fusion protein was cloned into the pJB02. This vector contains both the Ori P and Epstein Barr virus nuclear antigen (EBNA) components which are necessary for sustained, transient 25 expression in 293 EBNA cells. Further, this expression plasmid contains a puromycin resistance gene expressed from the CMV promoter as well as an ampicillin resistance gene. The gene of interest is also expressed from the CMV promoter.

30 The transfection mixture was prepared by mixing 73  $\mu$ l of the liposome transfection agent Fugene 6® (Roche Molecular Biochemicals, Cat. No. 1815-075) with 820  $\mu$ l Opti-Mem® (GibcoBRL Cat. No. 31985-062). G-CSF pJB02 DNA (12 $\mu$ g), prepared using a Qiagen plasmid maxiprep kit 35 (Qiagen, Cat. No. 12163), was then added to the mixture.

The mixture was incubated at room temperature for 15 minutes.

Cells were plated on 10 cm<sup>2</sup> plates in DMEM/F12 3:1 (GibcoBRL Cat. No. 93-0152DK) supplemented with 5% fetal bovine serum, 20mM HEPES, 2 mM L-glutamine, and 50 µg/mL Geneticin such that the plates were 60% to 80% confluent by the time of the transfection. Immediately before the transfection mixture was added to the plates, fresh media was added. The mixture was then added dropwise to cells with intermittent swirling. Plates were then incubated at 37°C in a 5% CO<sub>2</sub> atmosphere for 24 hours at which point the media was changed to hybritech medium without serum. The media containing a secreted form of a glycosylated G-CSF analog was then isolated 48 hours later.

15 2b: Expression in CHO cells:

The expression vector for expression in CHO-K1 cells is pEE14.1. This vector includes the glutamine synthetase gene which enables selection using methionine sulfoximine. This gene includes two poly A signals at the 3' end. G-CSF analogs are expressed from the CMV promoter which includes 5' untranslated sequences from the hCMV-MIE gene to enhance mRNA levels and translatability. The SV40 poly A signal is cloned 3' of the fusion protein DNA. The SV40 late promoter drives expression of GS minigene. This expression vector encoding the gene of interest is prepared for transfection using a QIAGEN Maxi Prep Kit (QIAGEN, Cat. No. 12362). The final DNA pellet (50-100 µg) is resuspended in 100 µl of basal formulation medium (GibcoBRL CD-CHO Medium without L-Glutamine, without thymidine, without hypoxanthine).

30 Before each transfection, CHO-K1 cells are counted and checked for viability. A volume equal to 1 x 10<sup>7</sup> cells is centrifuged and the cell pellet rinsed with basal formulation medium. The cells are centrifuged a second time and the final pellet resuspended in basal formulation medium (700 µl final volume).

The resuspended DNA and cells are then mixed together in a standard electroporation cuvette (Gene Pulsar Cuvette) used to support mammalian transfections, and placed on ice for five minutes. The cell/DNA mix is then electroporated in a BioRad Gene Pulsar device set at 300V/975  $\mu$ F and the cuvette placed back on ice for five minutes. The cell/DNA mixed is diluted into 20 ml of cell growth medium in a non-tissue culture treated T75 flask and incubated at 37°C / 5% CO<sub>2</sub> for 48-72 hours.

The cells are counted, checked for viability, and plated at various cell densities in selective medium in 96 well tissue culture plates and incubated at 37°C in a 5% CO<sub>2</sub> atmosphere. Selective medium is basal medium with 1X HT Supplement (GibcoBRL 100X HT Stock), 100  $\mu$ g/mL Dextran Sulfate (Sigma 100 mg/ml stock), 1X GS Supplements (JRH BioSciences 50X Stock) and 25  $\mu$ M MSX (Methionine Sulphoximine). The plates are monitored for colony formation and screened for glycosylated G-CSF analog production.

Example 3: Purification of Heterologous Fusion Proteins

Heterologous fusion proteins containing a hyperglycosylated G-CSF protein fused to albumin were harvested from host cells and then dialyzed against 20 mM Tris pH 7.4. An anion exchange column (1 ml Pharmacia HiTrap Q) was equilibrated with 20 mM Tris pH 7.4 and the dialyzed material loaded at 2 ml/min. The protein was eluted from the column using a linear gradient from 0 to 500 mM NaCl in 80 min at 1 ml/min and elution was monitored by UV absorbance at 280 nm. SDS-PAGE analysis was used to identify and pool fractions of interest. This pool was dialyzed against 25 mM sodium acetate (NaOAc) pH 5.0

A cation exchange column (1 ml Pharmacia HiTrap S column) was equilibrated with 25 mM NaOAc pH 5.0 and the dialysate was loaded at 1 ml/min. The protein was eluted

from the column using a linear gradient from 0 to 500 mM NaCl in 30 min. The fractions were immediately neutralized with 1 M Tris pH 8 to a final pH of 7. SDS-PAGE gels were used to identify and pool fractions of interest.

5        Heterologous fusion proteins containing a hyperglycosylated G-CSF protein fused to an Fc protein were harvested from host cells and dialyzed against 20 mM sodium phosphate pH 7.0. An affinity column (1 ml Pharmacia HiTrap Protein A or rProtein A) was equilibrated with 20 mM sodium phosphate pH 7.0 and the dialysate was loaded at 2 ml/min. 10        1 ml/min of 100 mM citric acid pH 3 was used to elute the protein. Fractions were immediately neutralized with 1M Tris pH 8 to pH 7 and peak fractions (determined by in-line OD280 monitoring) were further diluted with 20 mM sodium phosphate pH 7.0. 15        SDS-PAGE analysis was used to identify and pool fractions of interest.

WE CLAIM:

1. A heterologous fusion protein comprising a  
5 hyperglycosylated G-CSF analog fused to a polypeptide  
selected from the group consisting of
  - a) human albumin;
  - b) human albumin analogs; and
  - c) fragments of human albumin.
- 10 2. The heterologous fusion protein of claim 1, wherein the hyperglycosylated G-CSF analog is fused to the polypeptide via a peptide linker.
- 15 3. The heterologous fusion protein of Claim 2 wherein the peptide linker is selected from the group consisting of:
  - a) a glycine rich peptide;
  - b) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>n</sub> where n is 1, 2, 3, 4, or 5; and
  - 20 c) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>3</sub>.
- 25 4. The heterologous fusion protein of Claims 1, 2, or 3 wherein the hyperglycosylated G-CSF analog comprises the amino acid sequence of the formula I: [SEQ ID NO: 1]

1	5	10	15													
Thr	Pro	Leu	Gly	Pro	Ala	Ser	Ser	Leu	Pro	Gln	Ser	Phe	Leu	Leu	Lys	
20	20	25	30													
30	Xaa	Leu	Glu	Gln	Val	Arg	Lys	Ile	Gln	Gly	Asp	Gly	Ala	Ala	Leu	Gln
35	35	40	45													
45	Glu	Lys	Leu	Cys	Xaa	Xaa	Lys	Leu	Cys	His	Pro	Glu	Glu	Leu	Val	
50	50	55	60													
55	Leu	Leu	Gly	His	Ser	Leu	Gly	Ile	Xaa							
60	65	70	75	80												
65	Xaa	Xaa	Xaa	Xaa	Gln	Leu	Ala	Gly	Cys	Leu	Ser	Gln	Leu	His	Ser	
70	75	85	90	95												
75	Gly	Leu	Phe	Leu	Tyr	Gln	Gly	Leu	Leu	Gln	Ala	Leu	Xaa	Xaa	Ser	
80	85	100	105	110												
85	90	100	105	110												
95	95	115	120	125												
100	105	115	120	125												
110	115	120	125													
120	125	130	135	140												
130	135	130	135	140												
140	145	145	150	155	160											
150	155	150	155	160												

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Aln Ser Phe  
165 170  
Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro (I)

5 wherein:

Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;

Xaa at position 37 is Ala or Asn;

Xaa at position 38 is Thr, or any other amino acid except Pro;

10 Xaa at position 39 is Tyr, Thr, or Ser;

Xaa at position 57 is Pro or Val;

Xaa at position 58 is Trp or Asn;

Xaa at position 59 is Ala or any other amino acid except Pro;

15 Xaa at position 60 is Pro, Thr, Asn, or Ser,

Xaa at position 61 is Leu, or any other amino acid except Pro;

Xaa at position 62 is Ser or Thr;

Xaa at position 63 is Ser or Asn;

20 Xaa at position 64 is Cys or any other amino acid except Pro;

Xaa at position 65 is Pro, Ser, or Thr;

Xaa at position 66 is Ser or Thr;

Xaa at position 67 is Gln or Asn;

25 Xaa at position 68 is Ala or any other amino acid except Pro;

Xaa at position 69 is Leu, Thr, or Ser

Xaa at position 93 is Glu or Asn

Xaa at position 94 is Gly or any other amino acid except Pro;

30 Xaa at position 95 is Ile, Asn, Ser, or Thr;

Xaa at position 97 is Pro, Ser, Thr, or Asn;

Xaa at position 133 is Thr or Asn;

Xaa at position 134 is Gln or any other amino acid except Pro;

35 Pro;

Xaa at position 135 is Gly, Ser, or Thr

Xaa at position 141 is Ala or Asn;

Xaa at position 142 is Ser or any other amino acid except Pro; and

40 Xaa at position 143 is Ala, Ser, or Thr;

and wherein:

Xaa at positions 37, 38, and 39 constitute region 1;

Xaa at positions 58, 59, and 60 constitute region 2;

45 Xaa at positions 59, 60, and 61 constitute region 3;

Xaa at positions 60, 61, and 62 constitute region 4;

Xaa at positions 61, 62, and 63 constitute region 5;  
Xaa at positions 62, 63, and 64 constitute region 6;  
Xaa at positions 63, 64, and 65 constitute region 7;  
Xaa at positions 64, 65, and 66 constitute region 8;  
5 Xaa at positions 67, 68, and 69 constitute region 9;  
Xaa at positions 93, 94, and 95 constitute region 10;  
Xaa at positions 94, 95, and Ser at position 96  
constitute region 11;  
Xaa at positions 95, and 97, and Ser at position 96  
10 constitute region 12;  
Xaa at positions 133, 134, and 135 constitute  
region 13;  
Xaa at positions 141, 142, and 143 constitute  
region 14;  
15 and provided that at least one of regions 1 through 14  
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any  
amino acid except Pro and Xaa2 is Ser or Thr.

20 5. The heterologous fusion protein of Claim 4 wherein any  
two regions of regions 1 through 14 comprise the sequence  
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and  
Xaa2 is Ser or Thr.

25 6. The heterologous fusion protein of Claim 4 wherein any  
three regions of regions 1 through 14 comprise the sequence  
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and  
Xaa2 is Ser or Thr.

30 7. The heterologous fusion protein of Claim 4 wherein any  
four regions of regions 1 through 14 comprise the sequence  
Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and  
Xaa2 is Ser or Thr

8. The heterologous fusion protein of Claim 4 wherein the hyperglycosylated G-CSF analog is selected from the group consisting of:

- a) G-CSF[A37N, Y39T]
- 5 b) G-CSF[P57V, W58N, P60T]
- c) G-CSF[P60N, S62T]
- d) G-CSF[S63N, P65T]
- e) G-CSF[Q67N, L69T]
- f) G-CSF[E93N, I95T]
- 10 g) G-CSF[T133N, G135T]
- h) G-CSF[A141N, A143T]
- i) G-CSF[A37N, Y39T, P57V, W58N, P60T]
- j) G-CSF[A37N, Y39T, P60N, S62T]
- k) G-CSF[A37N, Y39T, S63N, P65T]
- 15 l) G-CSF[A37N, Y39T, Q67N, L69T]
- m) G-CSF[A37N, Y39T, E93N, I95T]
- n) G-CSF[A37N, Y39T, T133N, G135T]
- o) G-CSF[A37N, Y39T, A141N, A143T]
- p) G-CSF[A37N, Y39T, P57V, W58N, P60T, S63N, P65T]
- 20 q) G-CSF[A37N, Y39T, P57V, W58N, P60T, Q67N, L69T]
- r) G-CSF[A37N, Y39T, S63N, P65T, E93N, I95T]

9. The heterologous fusion protein of claim 8, wherein the hyperglycosylated G-CSF analog is G-CSF[A37N, Y39T, P57V, W58N, P60T, Q67N, L69T].

10. The heterologous fusion protein of claim 8, wherein the hyperglycosylated G-CSF analog is G-CSF[A37N, Y39T, S63N, P65T, E93N, I95T].

30 11. A heterologous fusion protein which is the product of the expression in a host cell of an exogenous DNA sequence which comprises a DNA sequence encoding a heterologous fusion protein of any one of Claims 1 through 10.

12. A polynucleotide encoding a heterologous fusion protein of any one of Claims 1 through 11.

13. A polynucleotide which comprises a DNA sequence

5 selected from the group consisting of:

a) SEQ ID NO:2

b) SEQ ID NO:3

c) SEQ ID NO:4

d) SEQ ID NO:5

10 e) SEQ ID NO:6

f) SEQ ID NO:7

g) SEQ ID NO:8

h) SEQ ID NO:9

i) SEQ ID NO:10

15 j) SEQ ID NO:11

k) SEQ ID NO:12

l) SEQ ID NO:13

m) SEQ ID NO:14

n) SEQ ID NO:15

20 o) SEQ ID NO:16,

fused in-frame to a DNA encoding a protein selected from the group consisting of:

a) human albumin,

b) human albumin analog; and

25 c) fragments of human albumin.

14. The polynucleotide of Claim 13, wherein the DNA fused in-frame comprises SEQ ID NO: 17.

30 15. The heterologous fusion protein of any one of Claims 1 through 11 wherein the polypeptide is human albumin.

16. The heterologous fusion protein of any one of Claims 1 through 11 wherein the polypeptide is an N-terminal fragment 35 of albumin.

17. A method for increasing neutrophil levels in a mammal comprising administering a therapeutically effective amount of the heterologous fusion protein of any one of Claims 1

40 through 11, 15 and 16.

18. The use of the heterologous fusion protein as claimed in any one of Claims 1 through 11, 15 and 16 for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

19. Use of a heterologous fusion protein of any one of Claims 1 through 11, 15, and 16 as a medicament.

10. 20. Use of a heterologous fusion protein of any one of Claims 1 through 11, 15, and 16 for the treatment of patients with insufficient circulating neutrophil levels.

15. 21. A pharmaceutical formulation adapted for the treatment of patients with insufficient neutrophil levels comprising a heterologous fusion protein of any one of Claims 1 through 11, 15, and 16.

20. 22. A heterologous fusion protein comprising a hyperglycosylated G-CSF analog fused to a polypeptide selected from the group consisting of:

25. a) the Fc portion of an immunoglobulin;  
b) an analog of the Fc portion of an immunoglobulin;  
and  
c) fragments of the Fc portion of an immunoglobulin.

30. 23. The heterologous fusion protein of Claim 22, wherein the hyperglycosylated G-CSF analog is fused to the polypeptide via a peptide linker.

35. 24. The heterologous fusion protein of the Claim 23 wherein the peptide linker is selected from the group consisting of:  
a) a glycine rich peptide;  
b) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>n</sub> where n is 1, 2, 3, 4, or 5; and  
c) a peptide having the sequence [Gly-Gly-Gly-Gly-Ser]<sub>3</sub>.

25. The heterologous fusion protein of Claims 22, 23 or 24, wherein the hyperglycosylated G-CSF analog comprises the amino acid sequence of the formula I: [SEQ ID NO: 1]

5	1	5	10	15
	Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Lys			
	20	25	30	
	Xaa Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln			
	35	40	45	
10	Glu Lys Leu Cys Xaa Xaa Xaa Lys Leu Cys His Pro Glu Glu Leu Val			
	50	55	60	
	Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa			
	65	70	75	80
15	Xaa Xaa Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser			
	85	90	95	
	Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Xaa Ser			
	100	105	110	
	Xaa Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp			
	115	120	125	
20	Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro			
	130	135	140	
	Ala Leu Gln Pro Xaa Xaa Xaa Ala Met Pro Ala Phe Xaa Xaa Xaa Phe			
	145	150	155	160
25	Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Aln Ser Phe			
	165	170		
	Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro (I)			

wherein:

30	Xaa at position 17 is Cys, Ala, Leu, Ser, or Glu;
35	Xaa at position 37 is Ala or Asn;
	Xaa at position 38 is Thr, or any other amino acid except Pro;
	Xaa at position 39 is Tyr, Thr, or Ser;
	Xaa at position 57 is Pro or Val;
40	Xaa at position 58 is Trp or Asn;
	Xaa at position 59 is Ala or any other amino acid except Pro;
	Xaa at position 60 is Pro, Thr, Asn, or Ser,
	Xaa at position 61 is Leu, or any other amino acid except Pro;
	Xaa at position 62 is Ser or Thr;
	Xaa at position 63 is Ser or Asn;
	Xaa at position 64 is Cys or any other amino acid except Pro;
45	Xaa at position 65 is Pro, Ser, or Thr;
	Xaa at position 66 is Ser or Thr;
	Xaa at position 67 is Gln or Asn;
	Xaa at position 68 is Ala or any other amino acid except Pro;
50	Xaa at position 69 is Leu, Thr, or Ser

Xaa at position 93 is Glu or Asn

Xaa at position 94 is Gly or any other amino acid except Pro;

Xaa at position 95 is Ile, Asn, Ser, or Thr;

5 Xaa at position 97 is Pro, Ser, Thr, or Asn;

Xaa at position 133 is Thr or Asn;

Xaa at position 134 is Gln or any other amino acid except Pro;

Xaa at position 135 is Gly, Ser, or Thr

10 Xaa at position 141 is Ala or Asn;

Xaa at position 142 is Ser or any other amino acid except Pro; and

Xaa at position 143 is Ala, Ser, or Thr;

15 and wherein:

Xaa at positions 37, 38, and 39 constitute region 1;

Xaa at positions 58, 59, and 60 constitute region 2;

Xaa at positions 59, 60, and 61 constitute region 3;

Xaa at positions 60, 61, and 62 constitute region 4;

20 Xaa at positions 61, 62, and 63 constitute region 5;

Xaa at positions 62, 63, and 64 constitute region 6;

Xaa at positions 63, 64, and 65 constitute region 7;

Xaa at positions 64, 65, and 66 constitute region 8;

Xaa at positions 67, 68, and 69 constitute region 9;

25 Xaa at positions 93, 94, and 95 constitute region 10;

Xaa at positions 94, 95, and Ser at position 96  
constitute region 11;

Xaa at positions 95, and 97, and Ser at position 96  
constitute region 12;

30 Xaa at positions 133, 134, and 135 constitute  
region 13;

Xaa at positions 141, 142, and 143 constitute  
region 14;

35 and provided that at least one of regions 1 through 14  
comprises the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any  
amino acid except Pro and Xaa2 is Ser or Thr.

26. The heterologous fusion protein of Claim 25 wherein any two regions of regions 1 through 14 comprise the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and 5 Xaa2 is Ser or Thr.

27. The heterologous fusion protein of Claim 25 wherein any three regions of regions 1 through 14 comprise the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and 10 Xaa2 is Ser or Thr.

28. The heterologous fusion protein of Claim 25 wherein any four regions of regions 1 through 14 comprise the sequence Asn Xaa1 Xaa2 wherein Xaa1 is any amino acid except Pro and 15 Xaa2 is Ser or Thr

29. The heterologous fusion protein of Claim 25 wherein the hyperglycosylated G-CSF analog is selected from the group consisting of:

20 a) G-CSF[A37N,Y39T]  
b) G-CSF[P57V,W58N,P60T]  
c) G-CSF[P60N,S62T]  
d) G-CSF[S63N,P65T]  
e) G-CSF[Q67N,L69T]  
25 f) G-CSF[E93N,I95T]  
g) G-CSF[T133N,G135T]  
h) G-CSF[A141N,A143T]  
i) G-CSF[A37N,Y39T,P57V,W58N,P60T]  
j) G-CSF[A37N,Y39T,P60N,S62T]  
30 k) G-CSF[A37N,Y39T,S63N,P65T]  
l) G-CSF[A37N,Y39T,Q67N,L69T]  
m) G-CSF[A37N,Y39T,E93N,I95T]  
n) G-CSF[A37N,Y39T,T133N,G135T]  
o) G-CSF[A37N,Y39T,A141N,A143T]  
35 p) G-CSF[A37N,Y39T,P57V,W58N,P60T,S63N,P65T]  
q) G-CSF[A37N,Y39T,P57V,W58N,P60T,Q67N,L69T]

r) G-CSF[A37N, Y39T, S63N, P65T, E93N, I95T]

30. The heterologous fusion protein of Claim 29 wherein the hyperglycosylated G-CSF analog is G-

5 CSF[A37N, Y39T, P57V, W58N, P60T, Q67N, L69T]

31. The heterologous fusion protein of Claim 29 wherein the hyperglycosylated G-CSF analog is G-  
CSF[A37N, Y39T, S63N, P65T, E93N, I95T]

10 32. The heterologous fusion protein of any one of Claims 22 through 31 wherein the polypeptide is the Fc portion of an Ig selected from the group consisting of: IgG1, IgG2, IgG3, IgG4, IgE, IgA, IgD, or IgM.

15 33. The heterologous fusion protein of Claim 32 wherein the polypeptide is the Fc portion of an Ig selected from the group consisting of: IgG1, IgG2, IgG3, and IgG4.

20 34. The heterologous fusion protein of Claim 33 wherein the polypeptide is the Fc portion of an IgG1 immunoglobulin.

35. The heterologous fusion protein of Claim 33 wherein the polypeptide is the Fc portion of an IgG4 immunoglobulin.

36. The heterologous fusion protein of any one of Claims 22 through 35 wherein the Fc portion is a human IgG protein.

30 37. The heterologous fusion protein of any one of Claims 22 through 36 wherein the Fc portion comprises hinge, CH2, and CH3 domains.

35 38. The heterologous fusion protein of Claim 34 wherein the polypeptide has the sequence of SEQ ID NO: 33.

39. A polynucleotide encoding a heterologous fusion protein of Claim 34, wherein the polynucleotide comprises SEQ ID NO: 22.

5 40. A polynucleotide encoding a heterologous fusion protein of any one of Claims 22 through 38.

41. A vector comprising the polynucleotide of Claim 12, 13, 14, 39 and 40.

10 42. A host cell comprising the vector of Claim 41.

43. A host cell expressing at least one heterologous fusion protein of any one of Claims 1 through 11, 15, 16, and 22 through 38.

44. The host cell of Claim 43 wherein said host cell is a CHO cell.

20 45. A process for producing a heterologous fusion protein comprising the steps of transcribing and translating a polynucleotide of Claim 40 under conditions wherein the heterologous fusion protein is expressed in detectable amounts.

25 46. A method for increasing neutrophil levels in a mammal comprising the administration of a therapeutically effective amount of the heterologous fusion protein of any one of Claims 27 through 36.

30 47. The use of the heterologous fusion protein as claimed in any one of Claims 22 through 29 for the manufacture of a medicament for the treatment of patients with insufficient circulating neutrophil levels.

35

48. Use of a heterologous fusion protein of any one of Claims 22 through 29 as a medicament.

49. Use of a heterologous fusion protein of any one of 5 Claims 22 through 29 for the treatment of patients with insufficient circulating neutrophil levels.

50. A pharmaceutical formulation adapted for the treatment 10 of patients with insufficient neutrophil levels comprising a heterologous fusion protein of any one of Claims 22 through 29.

51. A heterologous fusion protein as hereinbefore described 15 with reference to any one of the Examples.

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## Abstract

The present invention encompasses heterologous fusion proteins comprising a hyperglycosylated G-CSF analog fused to proteins such as albumin and the Fc portion of an immunoglobulin which act to extend the *in vivo* half-life of the protein compared to native G-CSF. These fusion proteins are particularly suited for the treatment of conditions treatable by stimulation of circulating neutrophils, such as after chemotherapy regimens or in chronic congenital neutropenia.

卷之三

Fig. 1

1/1

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 Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys  
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 Cys Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln  
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 Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ser  
 R3 R4 R5 R6 R7 R8  
 R2  
 R9 70 75 80  
Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser  
 85 90 R12  
 Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Xaa Ser Xaa  
 R10 R11  
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 Xaa Xaa Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp  
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Kuchibhotla, Uma

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Glu Lys Leu Cys Xaa Xaa Xaa Lys Leu Cys His Pro Glu Glu Leu Val  
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa  
50 55 60

Xaa Xaa Xaa Xaa Xaa Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser  
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Xaa Xaa Xaa Ser  
85 90 95

Xaa Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp  
100 105 110

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro  
115 120 125

Ala Leu Gln Pro Xaa Xaa Xaa Ala Met Pro Ala Phe Xaa Xaa Xaa Phe  
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&lt;223&gt; synthetic construct

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ccaactcttg tagaggtctc aagaaaccta ggaaaagtgg gcagcaaatg ttgtaaacat  
1320

cctgaagcaa aaagaatgcc ctgtgcagaa gactatctat ccgtggtcct gaaccagtta  
1380

tgtgtgttgc atgagaaaac gccagtaagt gacagagtca ccaaatgctg cacagaatcc  
1440

ttggtaaca ggcgaccatg ctttcagct ctggaagtgc atgaaacata cgttccaaa  
1500

gagtttaatg ctgaaacatt caccttccat gcagatataat gcacactttc tgagaaggag  
1560

agacaaatca agaaacaaac tgcacttgtt gagctcgtga aacacaagcc caaggcaaca  
1620

aaagagcaac tgaaagctgt tatggatgat ttgcagctt ttgttagagaa gtgctgcaag  
1680

gctgacgata aggagacctg ctttgcggag gagggtaaaa aacttgttgc tgcaagtcaa  
1740

gctgccttag gcttataatg ac  
1762

<210> 18

<211> 232

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 18

1 a Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro  
5 10 15

Ala Pro Glu Lys Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro  
20 25 30

Lys Asp Thr Lys Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val  
35 40 45

Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val  
50 55 60

Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln  
65 70 75 80

Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln  
85 90 95

Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala  
100 105 110

Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro  
115 120 125

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr  
130 135 140

Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser  
145 150 155 160

Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr  
165 170 175

Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr  
180 185 190

Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe  
195 200 205

Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys  
210 215 220

Ser Leu Ser Leu Ser Pro Gly Lys  
225 230

<210> 19

<211> 229

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 19

Glu Ser Lys Tyr Gly Pro Pro Cys Pro Pro Cys Pro Ala Pro Glu Phe  
1 5 10 15

Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr  
20 25 30

Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val  
35 40 45

Ser Gln Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp Gly Val  
50 55 60

Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe Asn Ser  
65 70 75 80

Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu  
85 90 95

Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu Pro Ser

100

105

110

Ser Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro  
115 120 125

Gln Val Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys Asn Gln  
130 135 140

Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala  
145 150 155 160

Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr  
165 170 175

Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Arg Leu  
180 185 190

Thr Val Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser Cys Ser  
195 200 205

Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser  
210 215 220

Leu Ser Leu Gly Lys  
225

<210> 20

<211> 585

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 20

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5p Ala His Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu  
5 10 15

Glu Asn Phe Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln  
20 25 30

Gln Cys Pro Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu  
35 40 45

Phe Ala Lys Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys  
50 55 60

Ser Leu His Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu  
65 70 75 80

Arg Glu Thr Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro  
85 90 95

Glu Arg Asn Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu  
100 105 110

Pro Arg Leu Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His  
115 120 125

Asp Asn Glu Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg  
130 135 140

Arg His Pro Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg  
145 150 155 160

Tyr Lys Ala Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala  
165 170 175

Cys Leu Leu Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser  
180 185 190

Ser Ala Lys Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu  
195 200 205

Arg Ala Phe Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro  
210 215 220

Lys Ala Glu Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys  
225 230 235 240

Val His Thr Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp  
245 250 255

Arg Ala Asp Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser  
260 265 270

Ser Lys Leu Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His  
275 280 285

Cys Ile Ala Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser  
290 295 300

Leu Ala Ala Asp Phe Val Glu Ser Lys Asp Val Cys Lys Asn Tyr Ala  
305 310 315 320

Glu Ala Lys Asp Val Phe Leu Gly Met Phe Leu Tyr Glu Tyr Ala Arg  
325 330 335

Arg His Pro Asp Tyr Ser Val Val Leu Leu Leu Arg Leu Ala Lys Thr  
340 345 350

Tyr Glu Thr Thr Leu Glu Lys Cys Cys Ala Ala Asp Pro His Glu  
355 360 365

Cys Tyr Ala Lys Val Phe Asp Glu Phe Lys Pro Leu Val Glu Glu Pro  
370 375 380

Gln Asn Leu Ile Lys Gln Asn Cys Glu Leu Phe Glu Asn Leu Gly Glu  
385 390 395 400

Tyr Lys Phe Gln Asn Ala Leu Leu Val Arg Tyr Thr Lys Lys Val Pro  
405 410 415

Gln Val Ser Thr Pro Thr Leu Val Glu Val Ser Arg Asn Leu Gly Lys  
420 425 430

Val Gly Ser Lys Cys Cys Lys His Pro Glu Ala Lys Arg Met Pro Cys  
435 440 445

Ala Glu Asp Tyr Leu Ser Val Val Leu Asn Gln Leu Cys Val Leu His  
450 455 460

Glu Lys Thr Pro Val Ser Asp Arg Val Thr Lys Cys Cys Thr Glu Ser  
465 470 475 480

Leu Val Asn Arg Arg Pro Cys Phe Ser Ala Leu Glu Val Asp Glu Thr  
485 490 495

Tyr Val Pro Lys Glu Phe Asn Ala Glu Thr Phe Thr Phe His Ala Asp  
500 505 510

Ile Cys Thr Leu Ser Glu Lys Glu Arg Gln Ile Lys Lys Gln Thr Ala  
515 520 525

Leu Val Glu Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu  
530 535 540

Lys Ala Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys  
545 550 555 560

Ala Asp Asp Lys Glu Thr Cys Phe Ala Glu Glu Gly Lys Lys Leu Val  
565 570 575

Ala Ala Ser Gln Ala Ala Leu Gly Leu  
580 585

<210> 21

<211> 703

<212> DNA

13> Artificial Sequence

<220>

<223> synthetic construct

<400> 21

gagcccaa at cttgtgac aa aactcacaca tgcccaccgt gcccagcacc tgaactcctg  
60

gggggaccgt cagtcttcc tttccccc aaacccaagg acaccctcat gatctcccg  
120

accctgagg tcacatgcgt ggtgggtggac gtgagccacg aagaccctga ggtcaagttc  
180

aactggtacg tggacggcgt ggaggtgc aatgccaaga caaagccgcg ggaggagcag  
240

tacaacagca cgtaccgtgt ggtcagcg tcaccgtcc tgcaccagga ctggctgaat  
300

ggcaaggagt acaagtgc a ggtctccaac aaagccctcc cagccccat cgagaaaacc  
360

atctccaa ag ccaaaggca gccccgagaa ccacaggtgt acaccctgcc cccatcccg  
420

gaggagatga ccaagaacca ggtcagcctg acctgcctgg tcaaaggctt ctatcccagc  
480

gacatcgccg tggagtggg a gacatggg cagccggaga acaactacaa gaccacgc  
540

cccggtctgg actccgacgg ctcccttcttc ctctatagca agtcaccgt ggacaagagc  
600

agggtggcagc aggggaacgt cttctcatgc tccgtatgc atgaggctt gcacaaccac  
660

tacacgcaga agagcctctc cctgtctccg ggtaaatgtat agt  
703

<210> 22

<211> 981

&lt;12&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; synthetic construct

&lt;400&gt; 22

tccaccaagg gcccattcggt cttcccgcta ggcgcctgct ccaggagcac ctccgagagc  
60acagccgccc tgggctgcct ggtcaaggac tacttccccg aaccggtgac ggtgtcggtgg  
120aactcaggcg ccctgaccag cggcggtgcac accttcccggt ctgtcctaca gtcctcagga  
180ctctactccc tcagcagcgt ggtgaccgtg ccctccagca gcttgggcac gaagacctac  
240acctgcaacg tagatcacaa gcccagcaac accaagggtgg acaagagagt tgagtccaaa  
300tatggtcccc catgcccacc ctgcccagca cctgagttcc tggggggacc atcagtcttc  
360ctgttccccca caaaacccaa ggacactctc atgatctccc ggaccctga ggtcacgtgc  
420gtgggtggtgg acgtgagcca ggaagacccc gaggtccagt tcaactggta cgtggatggc  
480gtggaggtgc ataatgccaa gacaaagccg cgggaggagc agttcaacag cacgtaccgt  
540gtggtcagcg tcctcaccgt cctgcaccag gactggctga acggcaagga gtacaagtgc  
600aaggctctcca acaaaggct cccgtcctcc atcgagaaaa ccatctccaa agccaaaggg  
660cagccccgag agccacaggt gtacaccctg ccccatccc aggaggagat gaccaagaac  
720caggtcagcc tgacctgcct ggtcaaaggc ttctacccca gcgacatcgac cgtggagtgg  
780

gagagcaatg ggcagccgga gaacaactac aagaccacgc ctcccggtgc ggactccgac

ggctccttct tcctctacag caggctaaccc gtggacaaga gcaggtggca ggagggaaat  
900

gtcttctcat gctccgtgat gcatgaggct ctgcacaacc actacacaca gaagagcctc  
960

tccctgtctc tggtaaatg a  
981

<210> 23

<211> 406

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 23

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys  
1 5 10 15

Ala Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln  
20 25 30

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val  
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys  
50 55 60

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser  
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser  
85 90 95

o Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp  
100 105 110

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro  
115 120 125

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe  
130 135 140

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe  
145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Glu Pro  
165 170 175

Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu  
180 185 190

Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp  
195 200 205

Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp  
210 215 220

Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly  
225 230 235 240

Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn  
245 250 255

Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp  
260 265 270

Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro  
275 280 285

Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu  
290 295 300

Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met Thr Lys Asn  
305 310 315 320

Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile  
325 330 335

Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr  
340 345 350

Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys  
355 360 365

Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys  
370 375 380

Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu  
385 390 395 400

Ser Leu Ser Pro Gly Lys  
405

<210> 24

<211> 403

<212> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 24

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys  
1 5 10 15

Ala Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln

20

25

30

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val  
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys  
50 55 60

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser  
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser  
85 90 95

Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp  
100 105 110

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro  
115 120 125

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe  
130 135 140

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe  
145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Glu Ser  
165 170 175

Lys Tyr Gly Pro Pro Cys Pro Ser Cys Pro Ala Pro Glu Phe Leu Gly  
180 185 190

Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met  
195 200 205

Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser Gln  
210 215 220

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5 Glu Asp Pro Glu Val Gln Phe Asn Trp Tyr Val Asp Gly Val Glu Val  
230 235 240

His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Phe Asn Ser Thr Tyr  
245 250 255

Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly  
260 265 270

Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Gly Leu Pro Ser Ser Ile  
275 280 285

Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val  
290 295 300

Tyr Thr Leu Pro Pro Ser Gln Glu Glu Met Thr Lys Asn Gln Val Ser  
305 310 315 320

Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu  
325 330 335

Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro  
340 345 350

Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Arg Leu Thr Val  
355 360 365

Asp Lys Ser Arg Trp Gln Glu Gly Asn Val Phe Ser Cys Ser Val Met  
370 375 380

His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser  
385 390 395 400

Leu Gly Lys

<210> 25

<211> 500

<12> PRT

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 25

Thr Pro Leu Gly Pro Ala Ser Ser Leu Pro Gln Ser Phe Leu Leu Lys  
1 5 10 15

Ala Leu Glu Gln Val Arg Lys Ile Gln Gly Asp Gly Ala Ala Leu Gln  
20 25 30

Glu Lys Leu Cys Ala Thr Tyr Lys Leu Cys His Pro Glu Glu Leu Val  
35 40 45

Leu Leu Gly His Ser Leu Gly Ile Pro Trp Ala Pro Leu Ser Ser Cys  
50 55 60

Pro Ser Gln Ala Leu Gln Leu Ala Gly Cys Leu Ser Gln Leu His Ser  
65 70 75 80

Gly Leu Phe Leu Tyr Gln Gly Leu Leu Gln Ala Leu Glu Gly Ile Ser  
85 90 95

Pro Glu Leu Gly Pro Thr Leu Asp Thr Leu Gln Leu Asp Val Ala Asp  
100 105 110

Phe Ala Thr Thr Ile Trp Gln Gln Met Glu Glu Leu Gly Met Ala Pro  
115 120 125

Ala Leu Gln Pro Thr Gln Gly Ala Met Pro Ala Phe Ala Ser Ala Phe  
130 135 140

Gln Arg Arg Ala Gly Gly Val Leu Val Ala Ser His Leu Gln Ser Phe  
145 150 155 160

Leu Glu Val Ser Tyr Arg Val Leu Arg His Leu Ala Gln Pro Gly Gly  
165 170 175

Gly Gly Ser Gly Gly Gly Ser Gly Gly Gly Ser Asp Ala His  
180 185 190

Lys Ser Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu Glu Asn Phe  
195 200 205

Lys Ala Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln Gln Cys Pro  
210 215 220

Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu Phe Ala Lys  
225 230 235 240

Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys Ser Leu His  
245 250 255

Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu Arg Glu Thr  
260 265 270

Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro Glu Arg Asn  
275 280 285

Glu Cys Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu Pro Arg Leu  
290 295 300

Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His Asp Asn Glu  
305 310 315 320

Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg Arg His Pro  
325 330 335

Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg Tyr Lys Ala  
340 345 350

Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala Cys Leu Leu

355

360

365

Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser Ser Ala Lys  
370 375 380

Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu Arg Ala Phe  
385 390 395 400

Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro Lys Ala Glu  
405 410 415

Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys Val His Thr  
420 425 430

Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp Arg Ala Asp  
435 440 445

Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser Ser Lys Leu  
450 455 460

Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His Cys Ile Ala  
465 470 475 480

Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser Leu Ala Ala  
485 490 495

Asp Phe Val Glu  
500

<210> 26

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<23> synthetic construct

<400> 26

gtaagcttgc gtcgacgcta gcggcgccgc gccatggccg gacctgccac ccagagcccc  
60

atgaagctg

69

<210> 27

<211> 61

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 27

ggggcagggga gctggctggg cccagtggag tggcttcctg cactgtccag agtgcactgt  
60

g

61

<210> 28

<211> 59

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 28

ggacagtgca ggaagccact ccactggcc cagccagctc cctgccccag agcttcctg  
59

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<210> 29

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 29

gaacctcgag gatcctcatt agggctgggc aaggtgcctt aagacgcggt acgacacaccc  
60

caggaagctc tg

72

<210> 30

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 30

gtaagcttgc gtcgacgcta gcggcgccgc gccatggcccg gacctgccac ccagagcccc  
60

atgaagctg

69

<210> 31

<211> 57

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 31

gctctaaggc cttgagcagg aagctctggg gcagggagct cgctgggccc agtggag  
57

<210> 32

<211> 53

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 32

gggcccagcg agctccctgc cccagagctt cctgctcaag gccttagagc aag  
53

<210> 33

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 33

gaacctcgag gatcctcatt agggctgggc aaggtgcctt aagacgcggt acgacacctc  
60

caggaagctc tg

72

<210> 34

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 34

gtaagcttgc gtcgacgcta gcggcgcgcc gccatggccg gacctgccac ccagagcccc  
60

atgaagctg

69

<210> 35

<211> 61

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 35

gtccgagcag cactagttcc tcgggggtggc acagcttgggt ggtgttacac agcttctcct  
60

g  
61

<210> 36

<211> 66

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 36

ggcgcagcgc tccaggagaa gctgtgtAAC accaccaAGC tgtGCCACCC cgaggaACTA  
60

gtgctg

66

<210> 37

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 37

gaacctcgag gatcctcatt agggctgggc aaggtgcctt aagacgcggt acgacacacctc  
60

caggaagctc tg  
72

<210> 38

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

23> synthetic construct

<400> 38  
gtaagcttgc gtcgacgcta gcggcgcgcc gccatggccg gacctgccac ccagagcccc  
60

atgaagctg  
69

<210> 39

<211> 61

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 39

ccccggcgct ggaaagcgct ggcgaaaggcc ggcatggcggtctggttggg ctgcaggcgca  
60

g  
61

<210> 40

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 40

ggcccttgcc ctgcagccca accagaccgc catgccggcc ttccgcagcg ctttccagcg  
60

P-15648.ST25.txt

<210> 41

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 41

gaacctcgag gatcctcatt agggctggc aaggtgcctt aagacgcggt acgacacac 60

caggaagctc tg

72

<210> 42

<211> 69

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 42

gtaagcttgc gtcgacgcta gcggcgccgc gccatggccg gacctgccac ccagagcccc 60

atgaagctg

69

<210> 43

<211> 68

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 43

gccccggcgct ggaaggtaga gttgaaggcc ggcatggcac cctgggtggg ctgaagagca  
60

ggggccat

68

<210> 44

<211> 74

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 44

ggaaatggcc cctgctttc agcccaccca gggtgccatg ccggccttca actctacctt  
60

ccagagccgg gcag

74

<210> 45

<211> 72

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

P-15648.ST25.txt

<200> 45

gAACCTCGAG GATCCTCATT AGGGCTGGGC AAGGTGCCTT AAGACGCGGT ACGACACCTC  
60

cAGGAAGCTC TG

72

<210> 46

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 46

gCTAGCGGCG CGCCACCATG  
20

<210> 47

<211> 33

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 47

gCTCAGGGTA GCGTTAACGA TGCCCAGAGA GTG  
33

<210> 48

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 48

gggcatcggtt aacgctaccc tgagcagctg  
30

<210> 49

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 49

gactcgagga tcctcattag ggctggg  
27

<210> 50

<211> 38

<212> DNA

<213> Artificial Sequence

<220>

<223> synthetic construct

<400> 50

gctagcggcg cgccaccatg gccggacctg ccacccag  
38

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